



## Colamine and tartaric acid based innovative antioxidant and anti-spoilage food additive for fish product industry

Aram R. Mikaelyan<sup>1,2</sup>, Tigran M. Soghomonyan<sup>3</sup>, Bella G. Babayan<sup>2,3,4</sup>, Samvel A. Bagdasaryan<sup>1,4</sup>, Nona L. Asatryan<sup>2</sup>,  
Anna M. Grigoryan<sup>1,2</sup>, Marina A. Melkumyan<sup>3</sup>

<sup>1</sup>Russian-Armenian (Slavonic) University, Yerevan, Armenia; <sup>2</sup>Agrobiotechnology Scientific Center, Branch of ANAU (Armenian National Agrarian University) Foundation, Ejmiatsin, Armenia; <sup>3</sup>Scientific and Production Center "Armbiotechnology", National Academy of Sciences of Republic of Armenia, Yerevan, Armenia; <sup>4</sup>Yerevan State University, Yerevan, Armenia

\***Corresponding author:** Anna M. Grigoryan, PhD, Senior researcher, Head of Chair of General and Pharmaceutical Chemistry, RAU, 123 Hovsep Emin St, 0051 Yerevan, Armenia.

**Submission date:** August 2<sup>nd</sup>, 2024; **Acceptance date:** September 27<sup>th</sup>; **Publication date:** October 7<sup>th</sup>, 2024

**Please cite this article as:** Mikaelyan A. R., Soghomonyan T. M., Babayan B G., Bagdasaryan S. A., Asatryan N. L., Grigoryan A. M., Melkumyan M. A. Colamine and Tartaric Acid Based Innovative Antioxidant and Anti-Spoilage Food Additive for Fish Product Industry. *Bioactive Compounds in Health and Disease* 2024; 7(10): 489-499

DOI: <https://www.doi.org/10.31989/bchd.v7i10.1429>

### ABSTRACT

**Background:** Fish farming is a vital sector of the global food industry, facing the challenge of developing efficient, chemical-free feeds for high-value fish species. To address this, innovative antimicrobials and antioxidants are crucial. This study explores the antioxidant and anti-spoilage properties of novel N-containing derivatives of natural L-tartaric acid (TA), a safe and widely used food additive.

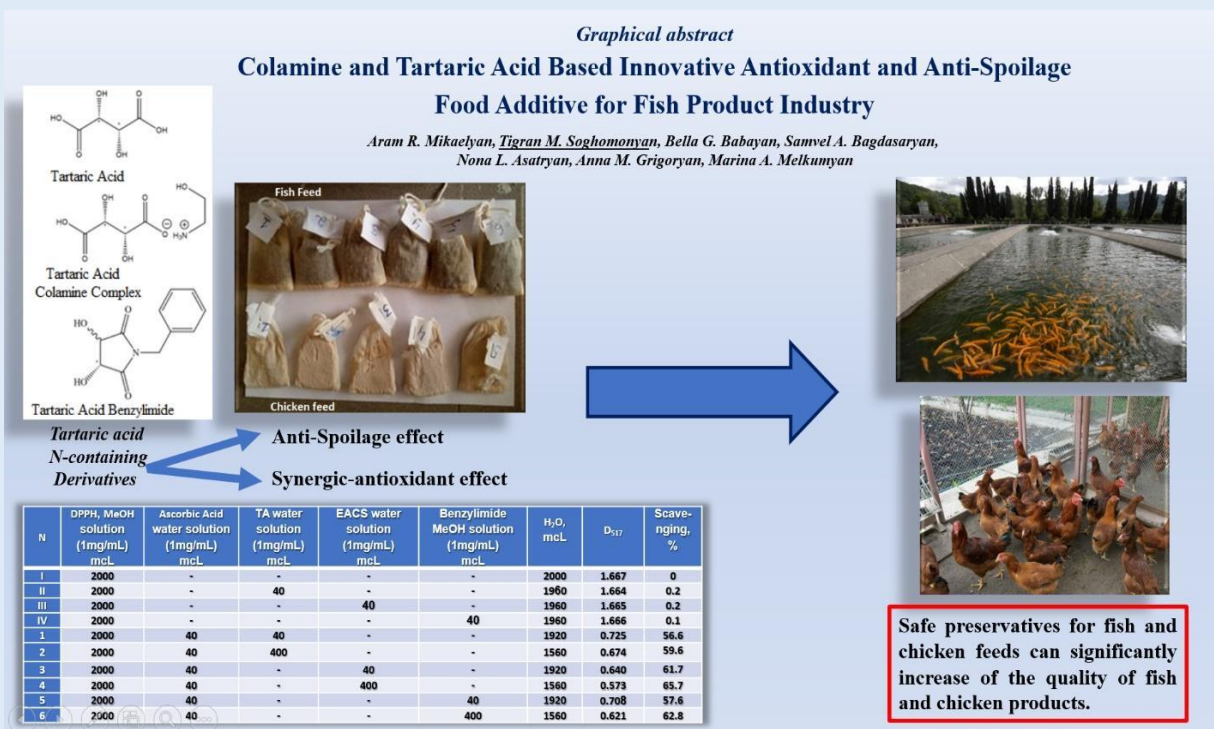
**Objectives:** Current research is devoted to L-tartrates' new derivatives' anti-spoilage and antioxidant potential as an innovative combined feed additive for fish farming.

**Results:** TA Cyclic imide obtaining modified technology was advised. According to the obtained results, TA itself didn't show antioxidant activity: nevertheless, in combination with new N-containing derivatives, it increased the antioxidant properties of L-ascorbic acid by about 20%. The studied TA amino-derivatives have demonstrated the emphasized activity against the fish-pathogenic *Pseudomonas*, *Streptococcus*, and other pathogenic microbes.

**Conclusions:** TA and tartrates, in combination with new N-containing derivatives generated based on the vital compound colamine, have demonstrated several synergic antioxidant activities. They can be recommended for potential application as a cheap and effective alternative to classical anti-spoilage and preservative agents for animal

feed production. Also, they showed the prolongation of feed expiration periods. It will significantly increase the quality of final fish products. A further detailed study of toxicological properties of amino derivatives is planned.

**Key words:** tartaric acid derivative, colamine, antimicrobial, antioxidant, anti-spoilage, feed additive



©FFC 2024. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (<http://creativecommons.org/licenses/by/4.0>)

## INTRODUCTION:

The fodder components of fish feeds, including the unsaturated fatty acids, amino acids, carbohydrates, hormones, vitamins, carotenoids, and other bioactive compounds (BAA) are exposed to intense oxidation during storage and usage. These unwanted processes result in the loss of valuable properties of fodder components and the formation and accumulation of toxic, dangerous products. It can significantly decrease the quality of final fish production, which can negatively affect human health [1-3].

Antioxidants, as bioactive compounds are very important for normal and pathological processes of all organisms, acting as inhibitors of oxidation processes [4-6]. This is related to the oxidative spoilage and microbial spoilage of food products [7]. They can have natural or synthetic origins. A generalized definition of antioxidants

was proposed in 1995: "as any substance, in the presence of low concentrations of which the oxidation process is significantly inhibited or prevented" [8-10]. According to this definition, not all the reducing agents involved in chemical reactions can be considered as the antioxidants. Antioxidants act as scavengers and neutralize the free radicals, resulting in oxidation processes being slowed down or completely inhibited. [11]. As a rule, free radicals have one or more free electrons, they can attach an atom or a whole molecule to them. Such phenomena in the body lead to oxidative stress, which causes a number of pathologies [12]. For this reason, antioxidants have a crucial role as a factor in health protection, and the balance maintained between the prooxidant and antioxidant systems in all living organisms. Scientific evidence suggests that antioxidants reduce the risk of

chronic diseases, including cancer and cardiovascular diseases. Additionally, they play a crucial role in aging control processes [13-14]. The global scientific community is currently conducting extensive research to develop novel antioxidant compounds with low toxicity, focusing on their potential applications in the production of food, animal feeds, and cosmetics [15].

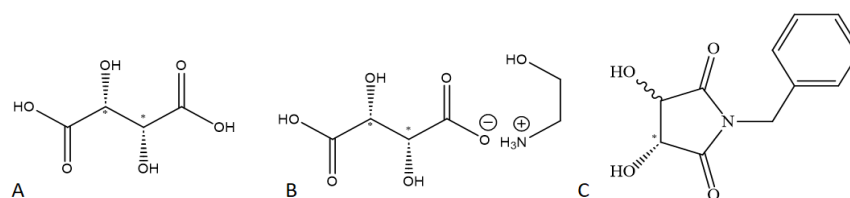
The application of synthetic and natural antioxidants is employed to inhibit oxidative processes and ensure the stability of food products. Natural compounds frequently display antioxidant properties, yet their effects can be ambiguous and context-dependent [16-19]. Among such compounds (bio-antioxidants) are: vitamins E, vitamin K, and vitamin B<sub>5</sub>, flavonoids (so-called vitamin P), lecithin and cephaline phosphatides, some phenols and polyphenols, serotonin, adrenaline, some amino acids and steroid hormones, benzoic acid, tannin, colamine (ethanolamine) and a few others. Excessive antioxidant intake can disrupt vitamin A synthesis from carotene, affecting intestinal absorption [20]. In aquaculture, antioxidant-supplemented fish feeds frequently incorporate synergistic preservative additives, specifically blends of organic acids (propionic, acetic, formic, benzoic, sorbic, lactic, and citric) and their salts, typically comprising up to 10% of the feed's total mass [21-23]. However, these synergistic-mixture additives exhibit a few technological complications, extrusion drawbacks, corrosion of feeding equipment, the likelihood of chemical burns, and sharp and unpleasant

odors [24-25].

For food supplements, the allowable daily intake (ADI) is a vital consideration. TA and its salts, being naturally derived compounds, offer advantages from this perspective [26]. Despite their natural origins, research has shown that TA and its salts exhibit modest antioxidant capabilities, highlighting the need for further investigation. Meanwhile, research has demonstrated that nutritional acids like citric acid, maleic acid, and acetic acid enhance the antioxidant activity of classical compounds (ascorbic acid, polyphenols, and carotenoids) when used together [27]. Specifically, the food industry utilizes TA and its compounds as harmless additives, such as TA (E 334), sodium tartrate (E 335ii), sodium hydrotartrate (E 336i), potassium tartrate (E 336ii), potassium hydrotartrate (E 336i), potassium sodium tartrate (E 337), and calcium tartrate (E 354). Their roles as acidity regulators, preservatives, and stabilizers make them suitable candidates for derivatization, enabling the creation of novel biomolecules with optimized functions [28-29]. Current research was devoted to the study of natural compounds and their functional derivatives application potential in feeds as anti-spoilage and antioxidant agents.

## MATERIALS AND METHODS:

**Synthesis of Tartaric acid derivatives:** TA and the studied derivatives of it are presented on Fig 1.



**Fig. 1.** Tartaric acid (TA) and its derivatives.

A – (*R,R*)-*L*-(+)-2,3-Dihydroxybutanedioic acid (TA); B – 2-hydroxyethan-1-aminium(2*R*,3*R*)-3-carboxy-2,3-dihydroxypropanoate (TA Colamine Complex); C – 1-benzyl-3,4-dihydroxy-pyrrolidine-2,5-dione (Benzylimide of TA or BI).

Additionally, amino derivatives of TA were synthesized using green chemistry technologies to modify TA [30]. The reaction process was controlled, and product formation was detected using a selective method of light absorption with ultraviolet rays (UV-254). Thin-layer chromatography (TLC) analyses were conducted on Silufol UV-254 plates [31]. Visualization was achieved by spraying with a 1% solution of ninhydrin to detect amines. Melting points were measured using a Fisher-Johns device.

Subsequently, the molecular characteristics of the target compound, 2-hydroxyethan-1-aminium(2R,3R)-3-carboxy-2,3-dihydroxypropanoate and 1-benzyl-3,4-dihydroxy-pyrrolidine-2,5-dione, were elucidated by Nuclear Magnetic Resonance (NMR) analysis. The NMR spectra were registered on a spectrometer Varian Mercury-300 at operating frequencies 300.077 MHz ( $^1\text{H}$ ), 75.46 MHz ( $^{13}\text{C}$ ), and chemical shifts were reported with respect to TMS (Tetramethylsilane) in  $\text{CDCl}_3$  (Deuterated chloroform). EACS and benzylimide were synthesized and investigated as new nitrogen-containing derivatives of natural tartaric acid. The synthesis modifications proposed by us were based on methods described in the literature [32].

The process of TA benzylimide (1-benzyl-3,4-dihydroxy-pyrrolidine-2,5-dione, also known as Benzylimide of Tartaric Acid) synthesis was carried out by following procedure: benzylamine (0.1 mol) and L-(+)-tartaric acid (0.1 mol) was refluxed for 8 – 10 h using a Dean-Stark apparatus. As the solvent *o*-xylene was used, at the temperature 183 – 185 °C. The reaction mixture was then cooled to ambient temperature, and the resulting crystalline product was separated by vacuum filtration. After washing portions of hexane, the filtrate was recrystallized from an acetonitrile-toluene mixture (v:v = 1:1). The final product was obtained as a pale solid powder with a 92% yield,  $m_p=200-202^\circ\text{C}$ .  $^1\text{H}$  NMR

spectrum in  $\text{CDCl}_3$ ,  $\delta$ , ppm (J, Hz): 4.38 (d, J = 4.6 Hz, 2H,  $-\text{CH}_2\text{Ph}$ ), 4.5 (d, J = 7.2 Hz, 2H,  $-\text{CH}-\text{CH}-$ ), 6.3 (d, J = 5.3 Hz, 2H,  $-\text{OH}$ ), 7.3-7.2 (m, 5H, ArH).

The process of 2-hydroxyethan-1-aminium-(2R,3R)-3-carboxy-2,3-dihydroxypropanoate (TA Colamine Complex) synthesis was carried out by the following procedure: TA (0.1 mol) was dissolved in  $\text{H}_2\text{O}$  (30 mL) and ethanolamine (0.1 mmol) was dissolved in EtOH (20 mL) before two solutions were mixed and incubated in a cold-water bath (at a temperature of  $-3^\circ\text{C} - 0^\circ\text{C}$ , created by the mixture of ice and NaCl) for 1 h. The precipitated salt was filtered off over suction and washed with diethyl ether affording TA colamine complex in the form of white crystals with a 95% yield,  $m_p=181-182^\circ\text{C}$ .  $^1\text{H}$  NMR spectrum in  $\text{DMSO}-d_6$  (Deuterated dimethyl sulfoxide) with addition of  $\text{CF}_3\text{COOH}$ ,  $\delta$ , ppm (J, Hz): 2.8 (br.s., 3H,  $\text{N}^+\text{H}_3-\text{CH}_2-$ ), 2.92 (m, 2H,  $\text{N}^+\text{H}_3-\text{CH}_2-$ ), 4.3 (s, 2H,  $\text{HO}-\text{CH}-\text{COO}^-$ ), 7.8 (br.s., 3H,  $-\text{OH}$ ).

**Antioxidant activity assessment:** Several methods assess antioxidant capacity, including ABTS (2,2-azinobis (3-ethylbenzothiazoline-6-sulfonic acid), FRAP (ferric reducing antioxidant power), CURPAC (cupric reducing antioxidant capacity), and DPPH (2,2-diphenyl-1-picrylhydrazyl) assays [33]. For this purpose, the spectroscopic method was applied, using DPPH 0.2 mM absolute methanol solution [34-36]. All the measurements we performed by the application of a spectrophotometer in the visible region at 517 nm, which corresponds to the absorption maximum of DPPH. The DPPH radical scavenging activity was calculated using the following equation (1):

$$\text{DPPH scavenging effect (\%)} = \{1 - (A_{\text{sample } 517 \text{ nm}} / A_{\text{control } 517 \text{ nm}})\} \times 100 \quad (1)$$

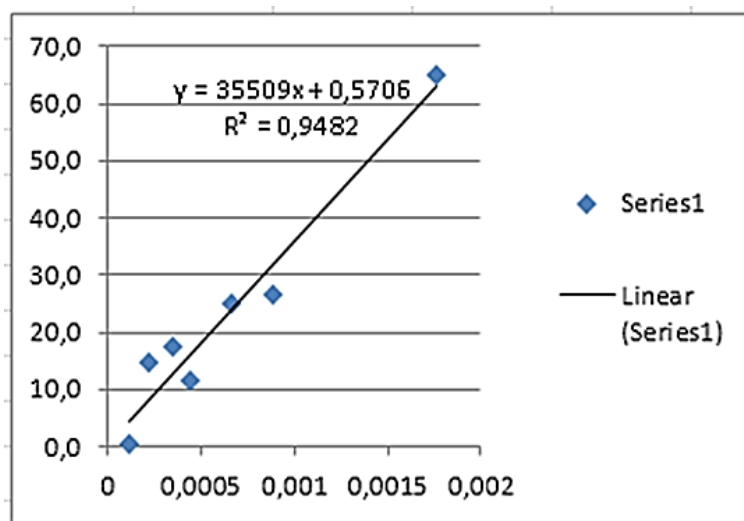
Where:  $A_{\text{control } 517 \text{ nm}}$  represents the absorbance value of the controlled reaction measured at 517 nm, using distilled water in place of test sample.  $A_{\text{sample } 517 \text{ nm}}$

represents the absorbance value of the test sample, measured at 517 nm, in presence of TA N-containing derivatives.

All the measurements were repeated five times and were performed at room temperature (23 °C). Photometric studies were conducted using a Thermo Scientific Multiskan GO spectrophotometer (517 nm). For the assessment of its value, the aqueous solutions were prepared by the consequent multiple dilutions of the initial 1 mg/mL ascorbic acid solution up to concentrations of 10, 20, 40, 80, and 500 µL, which were also diluted to a total volume of 2000 µL. The

mentioned solutions were mixed with 0.2 mM methanol solutions of DPPH. After the incubation of the mixture in the darkness for 30 min, the optical densities (D) of all the solutions were measured [37- 38].

Based on the collected data a grading curve was constructed using the least squares to a linear correlation in the concentration range 0 - 0.002 mg/ml ascorbic acid (see Fig. 1,  $y = 35509x + 0,5706$ , where  $y$  is the light absorption number, correlation coefficient  $R^2 = 0.9482$ ). The mean relative deviation was 2.23% ( $n = 5$ ). The value of  $EC_{50}$  for ascorbic acid was defined as 0.0014 mg/mL due to the constructed calibration curve.



**Figure 2.** The graphical presentation of DPPH neutralization dependence from the concentration of ascorbic acid in the titration solution.

### Anti-spoilage activity test

The anti-spoilage efficacy of TA benzylimide and colamine complex was assessed in two distinct feed formulations: fish feed and chicken feed. MODUS GRANUM Company (<https://modusgranum.am/>) supplied the feeds and their compositions. Fish feed preservation utilized "Vitasil", containing 40% formic acid, 30% acetic acid/peracetic acid, 15% sodium formate, 2% sodium benzoate, and 13% water (0.83% feed mass) (<https://agrovitex.ru/catalog/vitasil>). Chicken feed contained 0.5% sodium hydrosulfate. Tartaric acid

derivatives were added to the feed at 0.1% by weight. During batch preparation at the plant, individual samples were taken without antimicrobial additives, and then the final form of feed was applied.

The testing derivatives of TA were added to feeds without commercial antimicrobial additives using the same methodology, which is used for the feed production industry. To ensure uniform distribution, TA benzylimide was dissolved in DMSO, and TA colamine complex in water, as crystalline materials cannot be evenly dispersed in feed. 10 grams of feed were taken, and pre-prepared

solutions were added by spraying. Control samples included a negative control, comprising 10 mL water, TA, and DMSO, to assess the residual antimicrobial activity and moisture effects. The study incorporated a negative control sample, consisting of 10 mL water, TA, and DMSO, to evaluate baseline antimicrobial activity and moisture impact. For the positive control samples the commercial preservative "Vitasil" was applied [39].

The resulting masses were transferred into small air-permeable bags and stored in room conditions for 4 months. After 4 months, all samples were inoculated onto the surface of solid sterile nutrient agarised cultural

media (30 ml of meat peptone agar) in 90 mm Petri Dishes then were incubated for 72 h at 30-37 °C in aerobic conditions. After cultivation, the presence of bacteria was recorded by growth detection, using the method of counting of CFU (colony-forming unit) [40].

#### RESULTS:

The results of the studies of pure TA, its colamine derivative and TA benzylimide synergist antioxidant activity as feed additives for chicken and fish feeds are presented on Table 1.

**Table 1.** The comparison of the synergist antioxidant activity of tartaric acid (TA) derivatives.

N	DPPH	AA	TA	TA CC	TA B	W	D <sub>517</sub>	S, %
1	2000	-	-	-	-	2000	1.667	0
2	2000	-	40	-	-	1960	1.664	0.2
3	2000	-	-	40	-	1960	1.665	0.2
4	2000	-	-	-	40	1960	1.666	0.1
5	2000	40	40	-	-	1920	0.725	56.6
6	2000	40	400	-	-	1560	0.674	59.6
7	2000	40	-	40	-	1920	0.640	61.7
8	2000	40	-	400	-	1560	0.573	65.7
9	2000	40	-	-	40	1920	0.708	57.6
10	2000	40	-	-	400	1560	0.621	62.8

N – sample number (1-6); S – scavenging; TA – tartaric acid water solution (1 mg/mL) in mL; TA CC – tartaric acid colamine complex water solution (1 mg/mL) in mL; TA B – tartaric acid Benzylimide MeOH solution (1 mg/mL) in mL; AA – Ascorbic acid water solution (1 mg/mL) in mL; DPPH – DPPH, MeOH solution (1 mg/mL) in mL; W – water in mL.

The antioxidant activity measurements revealed that, in the absence of ascorbic acid, neither pure TA nor its derivatives (colamine complex and benzylimide) exhibited significant antioxidant properties. Specifically, after 30 minutes of interaction with DPPH, no substantial antioxidant effect was observed. The mean of the difference of the optical density value for the

observed solution was only 0.03 units. Organic acids are known to enhance ascorbic acid's antioxidant activity through significant synergistic effects [41]. As the next step of research, the anti-spoilage properties assessment was carried out. The results of anti-spoilage tests are presented in Fig. 3.



**Figure 3.** Anti-spoilage test of TA derivatives, as the feed additive for different feeds.

A: Fish feed, B: chicken feed. Samples: 1 – the industrial feed composition without preservative additives; 2 – the industrial feed with 0.83% “Vitasil” preservative; 3 – the industrial feed composition with the addition of 0.1% TA; 4 – the industrial feed composition with an addition of 0.1% TA colamine complex, 5 – the industrial feed composition with the addition of 0.1% TA benzylimide; 6 – the negative controls with the addition of 10 ml water and DMSO.

According to the obtained data, in all test samples, which contained the new synthetic derivatives of tartaric acid, the growth of bacteria was less pronounced. The maximum antimicrobial effect was expressed by TA benzylimide, in which the minimum intensity of microbial growth was demonstrated. Colamine complex of tartaric acid was characterized by a low antimicrobial effect, but it was more notable in comparison to samples with the industrial compositions with “Vitasil” or sodium hydrogen sulfate (or sodium bisulfate), preservatives application [42].

The study of the anti-spoilage effect of TA N-containing derivatives has demonstrated several antimicrobial effects against spoilage microbes. It was indicated that in the case of samples with N-containing TA

derivatives application, the titer of bacteria was  $2.2 \times 10^2$  –  $2.5 \times 10^2$  CFU for 1 g feed, stored within four months. For the positive control samples of the industrial feed composition with “Vitasil” and sodium hydrosulfate preservative the mean titer was  $3.7 \times 10^2$  –  $3.8 \times 10^2$ . Analysis of the negative control samples, which consisted of water and DMSO, yielded remarkably consistent mean titer values, with a narrow range of  $5.2 \times 10^2$  to  $5.4 \times 10^2$ .

The results indicate that the residual antimicrobial activity of DMSO does not substantially influence the growth of feed spoilage microorganisms. In contrast, humidity is a critical factor in their development and the subsequent deterioration of feed quality during prolonged storage.

## DISCUSSION:

The absence of ascorbic acid revealed no antioxidant activity in TA and its derivatives after 30 minutes of DPPH exposure, yet synergistic effects emerged when they were used together. The value of the optical density of the solution was changed by about 0.03 units. Research showed that TA colamine complex, TA benzylimide, and pure TA exhibit marked synergistic antioxidant properties when paired with ascorbic acid [43].

When applied at the same doses, or even at 10 times higher concentrations (400 mcL stock solution), the TA colamine complex showed the maximum synergy effect, exceeding pure TA by 5%. The TA benzylimide effect was 1% higher. Our findings suggest that TA derivatives outperform sodium sulfate, potentially enhancing fish and chicken feed quality by replacing sodium sulfate, which has several drawbacks (adverse effects on animal blood biochemistry, etc.) [44-46]. In accordance with that, TA N-containing derivatives application potentially might increase the level of food safety for humans.

Our comprehensive study demonstrates the promising prospects of researching TA's biological properties and synthetic derivatives, opening avenues for innovative applications. Tartaric acid's dual antioxidant and antimicrobial functionality presents opportunities for improved food and feed preservation methods and the creation of cutting-edge packaging solutions.

Studies on the antimicrobial properties of amino derivatives of tartaric acid demonstrated effectiveness against a variety of microorganisms, including fish pathogens, human opportunistic pathogens, and foodborne spoilage agents. The obtained data have proved the presence of synergic antioxidant activity in TA

and the colamine complex of TA. The creation of novel tartaric acid derivatives, especially those with amine functionality, represents a promising frontier in drug design and synthesis. The compounds synthesized demonstrate theoretical potential for varied biological effects, applicable to multiple areas of agriculture and healthcare.

**Abbreviations:** ABTS, 2,2-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid), allowable daily intake (ADI); BAA, bioactive compounds; CFU, colony-forming unit; CURPAC, cupric reducing antioxidant capacity; FRAP, ferric reducing antioxidant power; TA, L-tartaric acid; DPPH, 2,2-diphenyl-1-picrylhydrazine; MeOH, methanol.

**Author's Contributions:** All authors contributed to this study.

**Acknowledgments:** This work was supported by the Science Committee of RA (Research project № 23AA-1D018. We are also thankful to all the staff of "The Agrobiotechnology Scientific Center" Branch of ANAU Foundation, the Department of "Creation and Quality Control of Agricultural Preparations", SPC "Armbiotechnology" NAS RA Laboratory of Ecological Safety, The Chair of General and Pharmaceutical Chemistry of RAU, such as like to RAU staff for the organization of financial support for paper publication.

## REFERENCE

1. Tavares J, Martins A, Fidalgo LG, Lima V, Amaral RA, Pinto CA, Silva AM, Saraiva JA. Fresh Fish Degradation and Advances in Preservation Using Physical Emerging Technologies. *Foods*. 2021,10(4):780. DOI: <https://doi.org/10.3390/foods10040780>
2. Saini RV, Vaid P, Saini NK, Siwal SS, Gupta VK, Thakur VK, Saini AK. Recent Advancements in the Technologies Detecting Food Spoiling Agents. *J Funct Biomater*. 2021, 12(4):67. DOI: <https://doi.org/10.3390/ijfb12040067>



3. Snyder AB, Martin N, Wiedmann M. Microbial food spoilage: impact, causative agents and control strategies. *Nat Rev Microbiol.* 2024, 22(9):528-542.  
DOI: <https://doi.org/10.1038/s41579-024-01037-x>
4. Sotler R, Poljšak B, Dahmane R, Jukić T, Pavan Jukić D, Rotim C, Trebše P, Starc A. Prooxidant activities of antioxidants and their impact on health. *Acta Clin Croat.* 2019, 58(4):726-736.  
DOI: <https://doi.org/10.20471/acc.2019.58.04.20>
5. Maliar T, Maliarová M, Blažková M, Kunštek M, Uváčková Ľ, Viskupičová J, Purdešová A, Beňovič P. Simultaneously determined antioxidant and pro-oxidant activity of randomly selected plant secondary metabolites and plant extracts. *Molecules.* 2023; 28(19):6890. DOI: <https://doi.org/10.3390/molecules28196890>
6. Muscolo A, Mariateresa O, Giulio T, Mariateresa R. Oxidative stress: the role of antioxidant phytochemicals in the prevention and treatment of diseases. *Int J Mol Sci.* 2024, 25(6):3264.  
DOI: <https://doi.org/10.3390/ijms25063264>
7. Moreira D, Gullón B, Gullón P, Gomes A, Tavaría F. Bioactive packaging using antioxidant extracts for the prevention of microbial food-spoilage. *Food Funct.* 2016, 7(7):3273-82  
DOI: <https://doi.org/10.1039/C6FO00553E>
8. Awoyale W, Alamu EO, Ironi EA, Maziya-Dixon B, Menkir A. Impact of packaging materials and storage condition on retention of provitamin A carotenoids and xanthophylls in yellow-seeded maize flour. *Functional Foods in Health and Disease* 2018, 8(10): 472-487.  
DOI: <https://doi.org/10.31989/ffhd.v8i10.535>
9. Ma W, Li L. Trends and prospects in sustainable food packaging materials. *Foods.* 2024, 13(11):1744.  
DOI: <https://doi.org/10.3390/foods13111744>
10. Halliwell B, Murcia MA, Chirico S, Aruoma O. Free radicals and antioxidants in food and in vivo: what they do and how they work. *Crit Rev Food Sci Nutr* 1995, 35:7-20.  
DOI: <https://doi.org/10.1080/10408399509527682>
11. Hassanpour SH, Doroudi A. Review of the antioxidant potential of flavonoids as a subgroup of polyphenols and partial substitute for synthetic antioxidants. *Avicenna J Phytomed.* 2023; 13(4):354-376.  
DOI: <https://doi.org/10.22038/ajp.2023.21774>
12. Ghasemi S, Farokhpour F, Mortezaagholi B, Movahed E, Ghaedi A, Gargari MK, Khanzadeh M, Bazrgar A, Khanzadeh S. Systematic review and meta-analysis of oxidative stress and antioxidant markers in recurrent aphthous stomatitis. *BMC Oral Health.* 2023, 23(1):960.  
DOI: <https://doi.org/10.1186/s12903-023-03636-1>
13. Rusu ME, Fizeşan I, Vlase L, Popa DS. Antioxidants in age-related diseases and anti-aging strategies. *Antioxidants (Basel).* 2022, 11(10):1868.  
DOI: <https://doi.org/10.3390/antiox11101868>
14. Mrityunjaya M, Pavithra V, Neelam R, Janhavi P, Halami PM, Ravindra PV. Immune-boosting, antioxidant and anti-inflammatory food supplements targeting pathogenesis of COVID-19. *Front Immunol.* 2020, 11:570122  
DOI: <https://doi.org/10.3389/fimmu.2020.570122>
15. Poljsak B, Kovač V, Milisav I. Antioxidants, food processing and health. *antioxidants (Basel).* 2021, 10(3):433.  
DOI: <https://doi.org/10.3390/antiox10030433>
16. Mac Donald-Wicks L. K., Wood L. G., Garg M. L. Methodology for the determination of biological antioxidant capacity. 2006, *Sci Food Agric* 86: 2046-2056.  
DOI: <https://doi.org/10.1002/jsfa.2603>
17. Behera A, Dharmalingam Jothinathan MK, Rynthiang I, Saravanan S, Murugan R. Comparative antioxidant efficacy of green-synthesised selenium nanoparticles from pongamia pinnata, citrus sinensis, and acacia auriculiformis: an in vitro analysis. *Cureus.* 2024, 16(4):e58439.  
DOI: <https://doi.org/10.7759/cureus.58439>
18. Franca-Oliveira G, Hernández-Ledesma B, Martínez-Rodríguez AJ Bioactive peptides as an alternative treatment for Helicobacter pylori infection. *Bioactive Compounds in Health and Disease* 2024, 7(5): 245-264.  
DOI: <https://www.doi.org/10.31989/bchd.v7i5.1348>
19. El-Lateef HMA, El-Dabea T, Khalaf MM, Abu-Dief AM. Recent overview of potent antioxidant activity of coordination compounds. *Antioxidants (Basel).* 2023, 12(2):213.  
DOI: <https://doi.org/10.3390/antiox12020213>
20. Liu Q, Tang GY, Zhao CN, Gan RY, Li HB. Antioxidant activities, phenolic profiles, and organic acid contents of fruit vinegars. *Antioxidants (Basel).* 2019, 8(4):78.  
DOI: <https://doi.org/10.3390/antiox8040078>
21. Zhang B, Xia T, Duan W, Zhang Z, Li Y, Fang B, Xia M, Wang M. Effects of Organic Acids, Amino acids and phenolic compounds on antioxidant characteristic of zhenjiang aromatic vinegar. *Molecules.* 2019, 24(20):3799.  
DOI: <https://doi.org/10.3390/molecules24203799>
22. Garcia CS, da Rocha MJ, Presa MH, et al. Exploring the antioxidant potential of chalcogen-indolizines throughout *in vitro* assays. *Peer J.* 2024, 12:e17074.  
DOI: <https://doi.org/10.7717/peerj.17074>

23. Dos Santos FKF, Júnior AAMP, Filho ALN, et al. Graphene and Natural Products: A Review of antioxidant properties in graphene oxide reduction. *Int J Mol Sci.* 2024, 25(10):5182. DOI: <https://doi.org/10.3390/2Fijms25105182>
24. Wan J, Xi Q, Tang J, Liu T, Liu C, Li H, Gu X, Shen M, Zhang M, Fang J, Meng X. Effects of pelleted and extruded feed on growth performance, intestinal histology and microbiota of juvenile red swamp crayfish (*Procambarus clarkii*). *Animals (Basel).* 2022, 12(17):2252. DOI: <https://doi.org/10.3390/ani12172252>
25. Rossi S, Leso SM, Calovi M. Study of the Corrosion behavior of stainless steel in the food industry. *Materials (Basel).* 2024, 17(7):1617. DOI: <https://doi.org/10.3390/ma17071617>
26. Al-Tamimy SMA, Abbas Malik N, Taleb Dhiab A. The Effect of adding tartaric acid and salicylic acid on the chemical composition of eggs produced from lohman chickens. *Arch Razi Inst.* 2022, 77(6):2353-2357. DOI: <https://doi.org/10.22092/ari.2022.358734.2292>
27. Jantwal A, Durgapal S, Upadhyay J, Joshi T, Kumar A Chapter 4.17 - Tartaric acid. *Antioxidants Effects in Health the Bright and the Dark Side* 2022, 485-492, DOI: <https://doi.org/10.1016/B978-0-12-819096-8.00019-7>
28. Teshome E, Forsido SF, Rupasinghe HPV, Olika Keyata E. Potentials of natural preservatives to enhance food safety and shelf life: a review. *Scientific World Journal.* 2022; 2022: 9901018. DOI: <https://doi.org/10.1155/2022/9901018>
29. Coban HB. Organic acids as antimicrobial food agents: applications and microbial productions. *Bioprocess Biosyst. Eng.* 2020, 43(4):569-591. DOI: <https://doi.org/10.1007/s00449-019-02256-w>
30. Gómez-García M, Sol C, de Nova PJG, et al. Antimicrobial activity of a selection of organic acids, their salts, and essential oils against swine enteropathogenic bacteria. *Porcine Health Manag.* 2019, 5:32. DOI: <https://doi.org/10.1186/2Fs40813-019-0139-4>
31. Mikaelyan AR, Babayan BG, Grigoryan AL, Grigoryan AM, Asatryan NL, Melkumyan MA Tartaric acid new derivatives as prospective and safe alternative to antimicrobials for food products packing. *Functional Foods in Health and Disease* 2024, 14(1): 33-50. DOI: <https://www.doi.org/10.31989/ffhd.v14i1.1195>
32. Koli R, Mannur VS, Shetti PP. Robust high-performance thin-layer chromatography (HPTLC) method for stability assessment and simultaneous quantification of epigallocatechin-3-gallate and rosmarinic acid in lipid-based nanoparticles and biological matrices. *Phytochem Anal.* 2024. DOI: <https://doi.org/10.1002/pca.3360>
33. Gang Chen, Jiao Yan, Li Lili, Jie Zhang, Xuefan Gu, Hua Song Preparation and performance of amine-tartaric salt as potential clay swelling inhibitor. *Applied Clay Science* 138 (2017) 12-16. DOI: <http://dx.doi.org/10.1016/j.clay.2016.12.039>
34. Baliyan S, Mukherjee R, Priyadarshini A, Vibhuti A, Gupta A, Pandey RP, Chang CM. Determination of antioxidants by dpph radical scavenging activity and quantitative phytochemical analysis of ficus religiosa. *Molecules.* 2022, 27(4):1326. DOI: <https://doi.org/10.3390/molecules27041326>
35. Hossain TJ. Methods for screening and evaluation of antimicrobial activity: A review of protocols, advantages, and limitations. *Eur J Microbiol Immunol (Bp).* 2024, 14(2):97-115. DOI: <https://doi.org/10.1556/2F1886.2024.00035>
36. Boligon A, Machado M, Athayde M, Technical evaluation of antioxidant activity, *Medicinal chemistry* 2014, 47-55. DOI: <http://10.0.16.76/2161-0444.1000188>
37. Dhakal S, Aryal P, Aryal S. Bashyal D, Khadka D. Phytochemical and antioxidant studies of methanol and chloroform extract from leaves of *Azadirachta indica* A. Juss. in Tropical region of Nepal, *J. of Pharmacognosy and Phytotherapy*, 2016, 203-208. DOI: <https://doi.org/10.5897/JPP2016.0425>
38. Aghajanyan A, Hambarzumyan A, Minasyan E, Hovhannisyanyan G, Yeghyan K, Soghomonyan T, Avetisyan S, Sakanyan V, Tsaturyan A, Efficient isolation and characterization of functional melanin from various plant sources. *International Journal of Food Science & Technology*, 2024, 59 (6), 3545-3555. DOI: <https://doi.org/10.1111/ijfs.16506>
39. Roberto Lo Scalzo, Organic acids influence on DPPH\_ scavenging by ascorbic acid, *Italy 2008 Food Chemistry* 107, 40-43. DOI: <https://doi.org/10.1016/J.FOODCHEM.2007.07.070>
40. Wong ACH, Tian T, Tsoi JKH, Burrow MF, Matinlinna JP. Aspects of adhesion tests on resin-glass ceramic bonding. *Dent Mater.* 2017, 33(9):1045-1055. DOI: <https://doi.org/10.1016/j.dental.2017.06.013>
41. Heuser E, Becker K, Idelevich EA. Evaluation of an automated system for the counting of microbial colonies. *Microbiol Spectr.* 2023, 11(4):e0067323.

- DOI: <https://doi.org/10.1128/spectrum.00673-23>
42. Stolze N, Bader C, Henning C, Mastin J, Holmes AE, Sutlief AL. Automated image analysis with ImageJ of yeast colony forming units from cannabis flowers. *J Microbiol Methods*. 2019, 164:105681.  
DOI: <https://doi.org/10.1016/j.mimet.2019.105681>
43. Awed E, Sadek K, Soliman M, Khalil R Biochemical alterations in serum biomarkers of Nile tilapia (*Oreochromis niloticus*) exposed to sodium sulphate and *Spirulina platensis*. *Damanhour Journal of Veterinary Sciences*. 1: 4(1),1:1-6.  
DOI: <https://doi.org/10.21608/djvs.2020.23855.1009>
44. Liu B, Zhu J, Zhou Q, Yu D. Tolerance and safety evaluation of sodium sulfate: A subchronic study in laying hens. *Anim Nutr*. 2021, 7(2):576-586.
- DOI: <https://doi.org/10.1016/j.aninu.2020.08.009>
45. EFSA Panel on Food Contact Materials, Enzymes and Processing Aids (CEP), Lambré C, Barat Baviera JM, et al. Safety evaluation of the food enzyme laccase from the non-genetically modified *Trametes hirsuta* strain AE-OR. *EFSA J*. 2024, 22(5):e8778.  
DOI: <https://doi.org/10.2903/2Fi.efsa.2024.8778>
46. Baviera B, Bolognesi C, Cocconcelli PS, Crebelli R, Gott D.M., K. Grob, Lampi E, et al. Safety evaluation of the food enzyme laccase from the non-genetically modified *Trametes hirsuta* strain AE-OR. *EFSA J*. 2024, 22(5):e8778.  
DOI: <https://doi.org/10.2903/2Fi.efsa.2024.8778>