



Anti-inflammatory effects of hydrophobic constituents in the extract of the root cortex of *Paeonia suffruticosa*

Tatsuki Sato¹, Saki Shirako¹, Tetsuya Okuyama², Yukinobu Ikeya^{3,4}, and Mikio Nishizawa^{1*}

¹Department of Biomedical Sciences, College of Life Sciences, Ritsumeikan University, 1-1-1 Nojihigashi, Kusatsu, Shiga 525-8577, Japan; ²Department of Surgery, Kansai Medical University, 2-5-1 Shinmachi, Hirakata, Osaka 573-1010, Japan; ³Faculty of Pharmacy, Daiichi University of Pharmacy, 22-1 Tamagawa-cho, Minami-ku, Fukuoka 815-8511, Japan; and ⁴Ritsumeikan Asia-Japan Research Organization, 2-150 Iwakura-cho, Ibaraki, Osaka 567-8570, Japan

*Corresponding author: Professor Mikio Nishizawa, M.D., Ph.D., Department of Biomedical Sciences, College of Life Sciences, Ritsumeikan University; Kusatsu, Shiga 525-8577, Japan

Submission date: July 14th, 2022; Acceptance date: August 9th, 2022; Publication date: August 16th, 2022

Please cite this article as: Sato T., Shirako S., Okuyama T., Ikeya Y., Nishizawa M. Anti-inflammatory effects of hydrophobic constituents in the extract of the root cortex of *Paeonia suffruticosa*. *Bioactive Compounds in Health and Disease*. 2022; 5(8): 160-173. DOI:10.31989/bchd.v5i8.967

ABSTRACT

Background: The root cortex of *Paeonia suffruticosa* Andrews (Paeoniaceae), which is also called moutan bark, is known as *Botanpi* in Japan. This crude drug has been used in several Kampo formulas, such as *Daiobotanpito*, to treat menstrual disturbance and constipation by improving blood stasis and suppressing inflammation. However, the anti-inflammatory effect has not been well studied.

Objective: To clarify the anti-inflammatory effects of moutan bark, we isolated its constituents and investigated their activity.

Methods: Moutan bark was extracted and successively fractionated with ethyl acetate (EtOAc) and *n*-butanol. The constituents were analyzed using HPLC. The production of nitric oxide (NO), an inflammatory mediator, was measured in interleukin (IL)-1 β -treated rat hepatocytes to identify active fractions or constituents.

Results: The EtOAc-soluble fraction of moutan bark extract significantly inhibited NO production. Three hydrophobic constituents were isolated from this fraction and identified as paeonol (**1**), 1,2,3,4,6-pentagalloyl- β -D-glucose (β -PGG; **2**), and methyl gallate (**3**). Paeonol and paeoniflorin (**4**) were abundantly present in the EtOAc-soluble fraction and *n*-butanol-soluble fraction, respectively. The hydrophobic constituents suppressed NO production without exerting cytotoxicity and

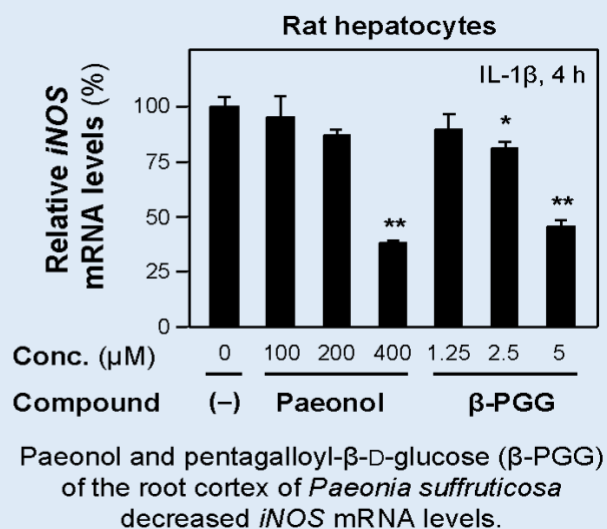
reduced the protein and mRNA levels of inducible nitric oxide synthase in IL-1 β -treated hepatocytes; β -PGG showed the highest potency. Furthermore, β -PGG, methyl gallate, and paeonol decreased the mRNA levels of tumor necrosis factor α and IL-1 receptor type 1, which are involved in inflammation.

Conclusion: These results suggest that hydrophobic constituents of moutan bark, such as β -PGG, methyl gallate, and paeonol, are involved in the anti-inflammatory effects of moutan bark.

Keywords: Galloylglucose, tannin, nitric oxide, Kampo medicine, cytokine.



Paeonia suffruticosa Andrews



©FFC 2022. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (<http://creativecommons.org/licenses/by/4.0/>)”

INTRODUCTION

Peony is a plant that is known for its beautiful flower and is part of the family Paeoniaceae, which includes *Paeonia suffruticosa* Andrews (formerly *Paeonia moutan* Sims). The root cortex of this species, also known as the moutan bark, is known as *Botanpi* in Japan [1]. This crude drug has been used in Japan and China [2] and included in several Japanese Kampo formulas. For example, *Daiobotanpito* improves blood stasis and suppresses inflammation in patients with menstrual disturbance and constipation. Moutan bark extract protects against oxidative injuries and inhibits the production of inflammatory mediators, such as nitric oxide (NO), interleukin (IL)-1 β , and tumor necrosis factor α (TNF- α), in lipopolysaccharide (LPS)-

induced RAW264.7 cells, which are a macrophage line [3]. NO plays a crucial role under physiological and pathophysiological conditions [4]. However, there have been few other reports on the anti-inflammatory effects of moutan bark.

The characteristic constituents of moutan bark are paeonol, paeoniflorin, and galloylglucose [2]. Paeonol, which is a major constituent, exerted anti-inflammatory effects and inhibited IL-1 β -induced inflammation in human chondrocytes and osteoarthritis model mice [5]. Paeoniflorin, a monoterpene glucoside, exerted anti-inflammatory and immunoregulatory effects on animal models of autoimmune diseases [6]. Galloylglucose is a simple hydrolysable tannin, and the major galloylglucose

in moutan bark is 1,2,3,4,6-pentagalloyl- β -D-glucose (β -PGG) [7], which is the β -anomer of an ester of glucose with five gallic acid molecules. Moutan bark exerts a variety of pharmacological effects, such as antioxidative, anti-inflammatory, hepatoprotective, and antitumor effects [2]. It is unclear which constituent primarily contributes to these effects of moutan bark.

Foreign pathogenic bacteria, such as Gram-negative bacteria, induce inflammatory responses, including NO production, through the release of LPS from the outer membrane of dead bacteria. LPS activates resident liver macrophages, such as Kupffer cells, to induce the expression of inducible nitric oxide synthase (iNOS), which synthesizes NO. Kupffer cells secrete IL-1 β , which binds to the IL-1 receptor type 1 (IL1R1) on hepatocytes to induce the expression of iNOS and proinflammatory cytokines through the IL-1 signaling pathway [8]. Primary cultured rat hepatocytes were used to assess anti-inflammatory activity because inhibiting IL-1 β -induced NO production is thought to be an anti-inflammatory marker. To date, the anti-inflammatory activity of many constituents of crude drugs, such as the rhizome of *Cnidium officinale* [9] and the bark of *Prunus jamasakura* [10], has been evaluated and compared using this *ex vivo* system.

Here, we purified pharmacologically active constituents from moutan bark and measured NO production in IL-1 β -stimulated hepatocytes. Then, the contents of the constituents were examined by HPLC. Finally, the anti-inflammatory effects of the constituents on the expression of *iNOS* and proinflammatory genes were examined.

MATERIALS AND METHODS

Plant materials: The root cortex of *Paeonia suffruticosa*

Andrews was collected from Anhui Province, China, and purchased from Tochimoto Tenkaido Co., Ltd. (Osaka, Japan). Dr. Yutaka Yamamoto (Tochimoto Tenkaido Co., Ltd.) authenticated it as *Botanpi*, according to the Japanese Pharmacopoeia [1]. The voucher specimen was deposited in the Ritsumeikan Herbarium of Pharmacognosy, Ritsumeikan University under RIN-PS-32.

Isolation of constituents from the bark of *Paeonia suffruticosa*:

The dried root cortex of *Paeonia suffruticosa* (370.0 g) was pulverized and extracted with methanol (MeOH) under reflux [11]. As shown in Figure 1, the extract (95.91 g) was suspended in water and sequentially partitioned with ethyl acetate (EtOAc) and *n*-butanol. These layers were concentrated to prepare EtOAc-soluble, *n*-butanol-soluble, and water-soluble fractions. The EtOAc-soluble fraction (Fraction A; 19.98 g), which suppressed NO production, was further purified by silica gel column chromatography [6.5 cm internal diameter (i.d.) \times 32 cm; Silica Gel 60, Nacalai Tesque, Inc., Kyoto, Japan] and was eluted with *n*-hexane:EtOAc (100:0 \rightarrow 0:100), acetone, and MeOH to yield 12 subfractions (A1 to A12). Subfractions A2 and A10 were further purified. Subfraction A2 (300 mg) was further fractionated using silica gel column chromatography (1.2 cm i.d. \times 9 cm; *n*-hexane:EtOAc = 80:20), resulting in one peak of Compound **1** (157 mg) as a white powder. Subfraction A10 was purified twice by ODS column chromatography (Cosmosil 75C18-OPN; 1.9 cm i.d. \times 30 cm; Nacalai Tesque, Inc.) and eluted with water:MeOH (100:0 \rightarrow 0:100) to yield Compound **2** (15.6 mg) as a brownish powder and Compound **3** (7.20 mg) as a brown powder.

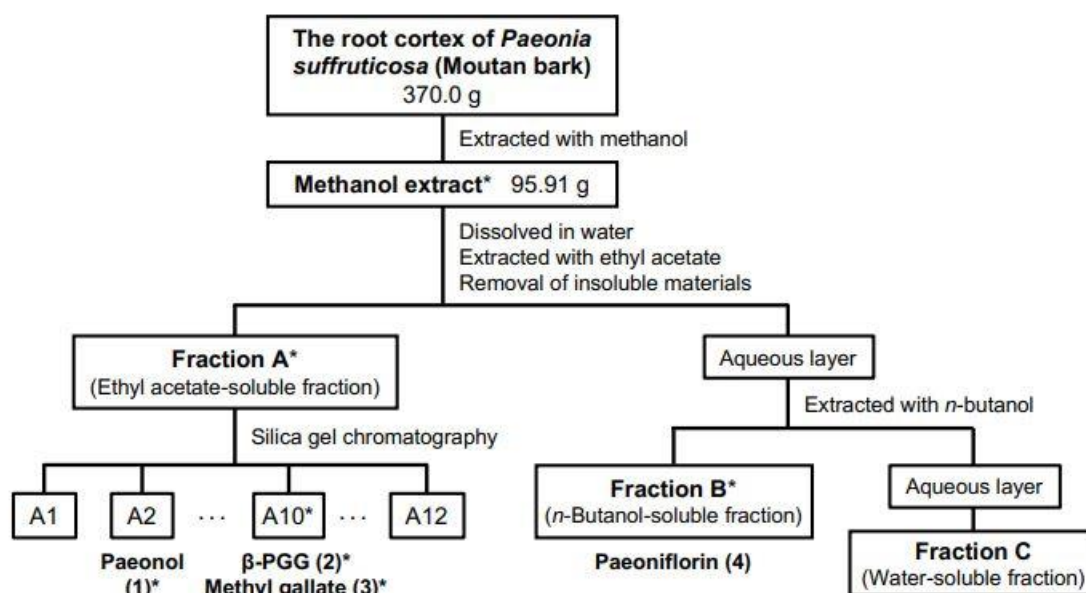


Figure 1. Purification of the constituents of the root cortex of *Paeonia suffruticosa*. A flowchart of the procedures used to fractionate the constituents of the root cortex of *Paeonia suffruticosa*. The yield of the extract (dried) was 25.9% of the plant material. The extract was separated into fractions A (hydrophobic fraction), B (amphipathic fraction), and C (hydrophilic fraction). Constituents included in the relevant subfraction are shown. * NO production was suppressed in hepatocytes in this study.

HPLC analysis of constituents: To measure the content of each constituent, an HPLC system equipped with an LC-20AD pump and an SPD-20A UV/VIS detector (Shimadzu Corporation, Kyoto, Japan) was used with a Cosmosil 5C18 AR-II column (4.6 mm i.d.×150 mm; Nacalai Tesque, Inc.). Paeonol, β-PGG, and methyl gallate were eluted at a flow rate of 1.0 mL/min by 0.1% (v/v) formic acid–MeOH (0 → 25 min; 80:20 → 0:100) and detected at a wavelength of 254 nm. Paeonol, β-PGG, and methyl gallate were accurately weighed and dissolved in MeOH to make stock solutions of 30 mM, 3 mM and 6 mM, respectively. Then, a series of paeonol (1.0, 2.0, 4.0 mM), β-PGG (0.0625, 0.125, 0.25 mM), and methyl gallate (0.05, 0.1, 0.2 mM) standard solutions were prepared by diluting the stock solutions to make calibration curves. A calibration curve for each standard compound was prepared by plotting peak areas (y) against a series of injection amounts (x μ g). The calibration equation and correlation coefficient of the three standard compounds

were as follows: paeonol, $y = 3878582x - 254175$ ($R^2 = 0.9994$); β-PGG, $y = 16418638x - 40279$ ($R^2 = 0.9999$); and methyl gallate, $y = 6733859x - 19490$ ($R^2 = 0.9999$). Fraction A was accurately weighed and dissolved in MeOH to make a 1.0 mg/mL sample solution. The peak areas of paeonol, β-PGG, and methyl gallate in the sample solution were fit to the calibration curves, and the amounts of each compound in 20 μ L of the sample solution were calculated. The amounts of paeonol, β-PGG, and methyl gallate in 20 μ L of the sample solution (20 μ g of Fraction A) were calculated to be 6.5 μ g, 2.3 μ g, and 0.29 μ g, respectively; therefore, the contents of these compounds in fraction A were 32.5%, 11.5%, and 1.47%, respectively.

To measure the paeoniflorin content, HPLC was performed. Paeoniflorin was eluted at a flow rate of 1.0 mL/min by 0.1% (v/v) formic acid–MeOH (72:28) and detected at a wavelength of 232 nm. Paeoniflorin (Tokyo Chemical Industry Co., Ltd., Tokyo, Japan) was dissolved

in MeOH, and a series of paeoniflorin standard solutions were prepared by diluting the stock solution to make calibration curves. The calibration equation and correlation coefficient of the paeoniflorin standard were $y = 360047900x - 1752.59$ ($R^2 = 0.9998$). Fraction B was dissolved in MeOH to prepare a 1.0 mg/mL sample solution and diluted to 0.5 mg/mL with 10% (v/v) acetonitrile containing 0.1% (v/v) formic acid. The peak areas of paeoniflorin in the sample solution were fit to the calibration curves. Because the amount of paeoniflorin in 20 μ L of the sample solution (10 μ g of Fraction B) was calculated to be 1.49 μ g, the paeoniflorin content in fraction B was 14.9%.

Nuclear magnetic resonance (NMR), polarimetry, and mass spectrometry: A JNM-ECS400 NMR spectrometer (JEOL Ltd., Akishima, Tokyo, Japan) operated at 400 MHz (^1H) and 100 MHz (^{13}C) was used to record NMR spectra. Deuterated chloroform (CDCl_3), methanol (CD_3OD) and tetramethylsilane (an internal standard) were purchased from Eurisotop (Saint-Aubin, France). A DIP-1000 polarimeter (JASCO Corporation, Hachioji, Tokyo, Japan) was used to measure the optical rotations. Spectra using fast atom bombardment mass spectrometry (FAB-MS) were obtained with a JMS-700 MStation mass spectrometer (JEOL Ltd.).

Animal experiments: Specific pathogen-free male Wistar rats (5–6 weeks old; Charles River Laboratories Japan, Inc., Yokohama, Japan) were housed at 21–23 $^{\circ}\text{C}$ with a 12 h light-dark cycle. The rats were fed a γ -ray-irradiated CRF-1 diet (Charles River Laboratories Japan, Inc.) and water *ad libitum*. All animal care and experimental procedures were performed in accordance with the laws and guidelines of the Japanese government and were approved by the Animal Care Committee of Ritsumeikan University, Biwako-Kusatsu Campus (No. BKC2020-045).

Primary cultured rat hepatocytes: After at least one week of acclimatization, hepatocytes were isolated from the livers of Wistar rats [12]. Briefly, the liver was perfused with collagenase (FUJIFILM Wako Pure Chemical Corporation, Osaka, Japan), and the dispersed cells were centrifuged, resuspended, and seeded at 1.2×10^6 cells per 35 mm diameter dish. The cells were incubated at 37 $^{\circ}\text{C}$ for 2 h, and then the medium was replaced (Day 0). The hepatocytes were further incubated at 37 $^{\circ}\text{C}$ overnight for subsequent assays.

NO production and lactate dehydrogenase (LDH) activity: On Day 1, the hepatocytes were incubated with each fraction or compound with/without 1 nM IL-1 β for 8 h. The Griess method [13–14] was used to measure nitrite, which is a stable metabolite of NO, in the medium in triplicate. The NO concentration in the medium alone was set as 0%, and the NO concentration in medium containing IL-1 β was set as 100%. The IC_{50} values of nitrite were calculated for at least three different concentrations of a fraction or a compound, unless it showed cytotoxicity [14]. Loxoprofen sodium (Kolon Life Science, Incheon, South Korea) was used as a positive control [14]. Hepatocyte cytotoxicity was assessed by measuring LDH activity in the culture medium using an LDH Cytotoxicity Detection Kit (Takara Bio Inc., Otsu, Japan). To evaluate direct NO-quenching activity, each compound was added to medium containing 25 μM NaNO_2 and incubated at 37 $^{\circ}\text{C}$ for 1.5 h [15]. The medium was mixed with Griess reagent [13–14] and incubated at 20–23 $^{\circ}\text{C}$ for 5 min, and the absorbance at 540 nm was measured to determine the decrease in nitrite induced by the compound.

Western blot analysis: Hepatocytes were treated with 1 nM IL-1 β and a compound for 8 h. Cell lysates (20 μg protein/lane) were resolved by a 10% sodium dodecyl sulfate–polyacrylamide gel and transferred onto a membrane [16]. Blocking with 5% Difco skim milk (BD

Biosciences, San Jose, CA, USA) and immunostaining were carried out using antibodies against iNOS (BD Biosciences) and β -tubulin (Cell Signaling Technology Inc., Danvers, MA, USA; internal control) and horseradish peroxidase-conjugated anti-immunoglobulin Fc antibodies (Cell Signaling Technology Inc.). ECL Western Blotting Detection Reagents (Cytiva, Tokyo, Japan) were used to visualize the proteins, and chemiluminescence was then detected using an Amersham Imager 600 (Cytiva).

Quantitative reverse transcription-polymerase chain reaction (RT-PCR): Sepasol I Super G solution (Nacalai Tesque, Inc.) was used to extract total RNA from hepatocytes, which was then purified [17–18]. The resultant RNA was reverse-transcribed to cDNA, which was then amplified by PCR with primers [17–18]. Real-time PCR was performed with SYBR Green I and a Thermal Cycler Dice Real Time System (Takara Bio Inc.), and the mRNA levels were measured in triplicate. After calculation by the $\Delta\Delta C_t$ method, C_t values were normalized to elongation factor 1 α (EF) mRNA [9,17–18]. The mRNA levels that were normalized to the total RNA from hepatocytes in the presence of IL-1 β alone were set as 100%.

Statistical analysis: The results are representative of at least three independent experiments that yielded similar findings. The values are presented as the mean \pm standard deviation (SD). The differences were analyzed using Student's t test followed by the Bonferroni correction. Significance was set at $P < 0.05$ and $P < 0.01$.

RESULTS

Hydrophobic constituents in the EtOAc-soluble fraction of moutan bark extract:

The root cortex of *Paeonia suffruticosa* (moutan bark) was extracted with MeOH and was successively fractionated with EtOAc and *n*-butanol based on hydrophobicity (Figure 1). NO production in hepatocytes was monitored, and the results demonstrated that the EtOAc-soluble fraction (Fraction A) and *n*-butanol-soluble fraction (Fraction B) of moutan bark extract decreased IL-1 β -induced NO production, whereas the water-soluble fraction (Fraction C) did not affect NO production (data not shown). The LDH assay showed that cytotoxicity was not observed at the concentrations used (data not shown). As summarized in Table 1, Fraction A gave the highest IC₅₀ value for NO production. Therefore, the hydrophobic constituents with NO-suppressing activity in this fraction were further purified.

Table 1. Fractionation of the extract from the root cortex of *Paeonia suffruticosa* (moutan bark) and its effects on NO production.

Fraction	Yield [%] ^a	IC ₅₀ [μ g/mL] ^b
MeOH extract	100	36.2 \pm 7.64
EtOAc-soluble fraction (Fraction A)	22.2	8.97 \pm 0.06
<i>n</i> -Butanol-soluble fraction (Fraction B)	9.93	141 \pm 23.5
Water-soluble fraction (Fraction C)	67.9	NA

^aThe percentage was calculated by determining the weight of each fraction divided by the sum of three fractions. ^bThe half-maximal inhibitory concentration of NO production in IL-1 β -stimulated hepatocytes (mean \pm SD). At least three experiments were performed to determine these values. NA, not applied due to low activity.

The constituents of Fraction A were purified by silica gel chromatography. Three hydrophobic constituents were purified from the subfractions that suppressed NO production, as described in the Materials and Methods. The constituents identified were paeonol (Compound **1**), 1,2,3,4,6-pentagalloyl- β -D-glucose (β -PGG; **2**), and methyl gallate (**3**). The values observed by the analyses are shown below. The chemical structures are shown in Figure 2.

Compound **1**: melting point (mp): 46–48 °C; $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ : 12.8 (1H, s, 2-OH), 7.63 (1H, d, $J = 8.5$ Hz, H-6), 6.46 (1H, d, $J = 2.4$ Hz, H-3), 6.43 (1H, dd, $J = 7.3$, 2.4 Hz, H-5), 3.84 (3H, s, 4-OMe), 2.56 (3H, s, H-8). $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 202.6 (C-7), 166.1 (C-4), 165.3 (C-2), 132.3 (C-6), 113.9 (C-1), 107.7 (C-5), 100.8 (C-3), 55.6 (4-OMe), 26.3 (C-8). The ^1H NMR and ^{13}C NMR spectra of **1** were identical to the previously reported ^1H NMR and ^{13}C NMR spectra of paeonol [19]. The mp of **1** was similar to previously reported mp values (46–49 °C) [20].

Compound **2**: $[\alpha]_{20}^D +31.0$ (c 0.338, MeOH). FAB-MS m/z 963 $[\text{M} + \text{Na}]^+$; HR-FAB-MS m/z 963.1076 $[\text{M} + \text{Na}]^+$ (calculated for $\text{C}_{41}\text{H}_{32}\text{O}_{26}\text{Na}$; 963.1080); ^1H NMR (400 MHz, CD_3OD) δ : 7.11 (2H, s, H-2'''' and H-6'''''), 7.05 (2H,

s, H-2'''' and H-6'''''), 6.98 (2H, s, H-2' and H-6'), 6.95 (2H, s, H-2''' and H-6'''), 6.90 (2H, s, H-2'' and H-6''), 6.24 (1H, d, $J = 8.6$ Hz, H-1), 5.91 (1H, t, $J = 9.6$ Hz, H-3), 5.62 (1H, t, $J = 9.6$ Hz, H-4), 5.59 (1H, dd, $J = 9.6$, 8.6 Hz, H-2), 4.52 (1H, brd, $J = 10.8$ Hz, H_a-6), 4.42 (1H, m, H-5), 4.39 (1H, d, $J = 10.8$, 4.5 Hz, H_b-6). $^{13}\text{C-NMR}$ (100 MHz, CD_3OD) δ : 168.0, 167.3, 167.1, 167.0, 166.2 (C-7'/C-7''/C-7'''/C-7''''/C-7'''''), 146.6, 146.5, 146.5, 146.4, 146.3 (C-3' and C-5'/C-3'' and C-5''/C-3''' and C-5'''/C-3'''' and C-5''''/C-3'''''), 140.8, 140.4, 140.3, 140.2, 140.0 (C-4'/C-4''/C-4'''/C-4''''/C-4'''''), 121.1, 120.4, 120.3, 120.2, 119.8 (C-1'/C-1''/C-1'''/C-1''''/C-1'''''), 110.6, 110.5, 110.4, 110.4, 110.4, 110.4 (C-2' and C-6'/C-2'' and C-6''/C-2''' and C-6'''/C-2'''' and C-6''''/C-2'''''), 93.8 (C-1), 74.4 (C-5), 74.1 (C-3), 72.2 (C-2), 69.8 (C-4), 63.1 (C-6). The ^1H and ^{13}C NMR spectra of **2** were identical to the previously published ^1H and ^{13}C NMR spectra of 1,2,3,4,6-pentagalloyl- β -D-glucose (PGG) [21].

Compound **3**: $^1\text{H-NMR}$ (400 MHz, CD_3OD) δ : 7.04 (2H, s, H-2/H-6), 3.81 (3H, s, H-8). ^{13}C NMR (100 MHz, CD_3OD) δ : 167.8 (C-7), 145.3 (C-3 and C-5), 138.6 (C-4), 120.2 (C-1), 108.8 (C-2 and C-6), 51.1 (C-8). The ^1H and ^{13}C NMR spectra of **3** were identical to the previously published ^1H and ^{13}C NMR spectra of methyl gallate [22].

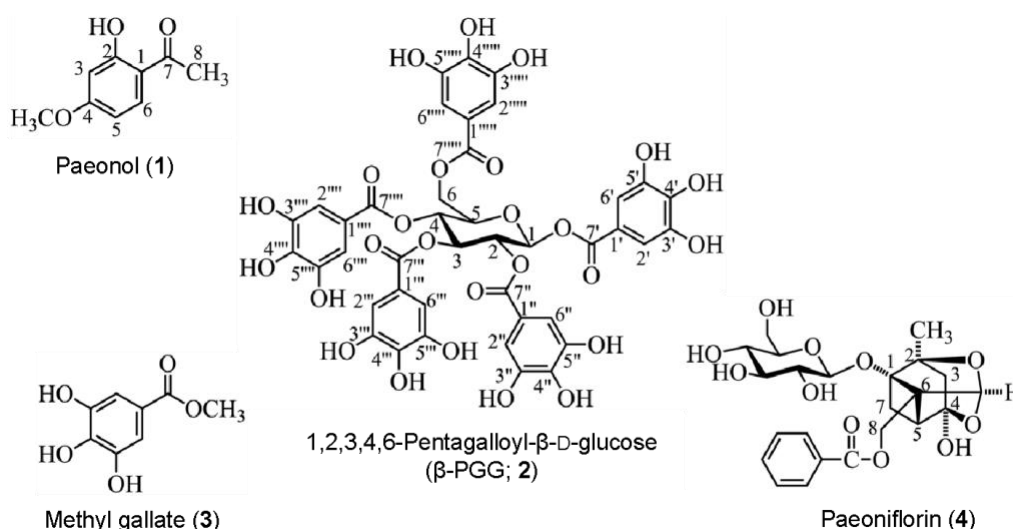


Figure 2. Constituents in moutan bark. The chemical structures of the constituents isolated from Fraction A of moutan bark extract (Compounds **1** to **3**) were determined by the spectral data. The chemical structure of paeoniflorin (Compound **4**) in fraction B is also shown.

The content of the constituents in moutan bark: The HPLC results indicated three independent peaks in the chromatogram of Fraction A (Figure 3), and then the content of each constituent was estimated. The content of paeonol (**1**) was the highest compared with the

content of the other constituents in Fraction A (Table 2). Paeonol content in moutan bark extract was calculated, and paeonol was confirmed to be a major constituent. In addition, HPLC analysis demonstrated that paeoniflorin (**4**) was abundantly present in Fraction B (Table 2).

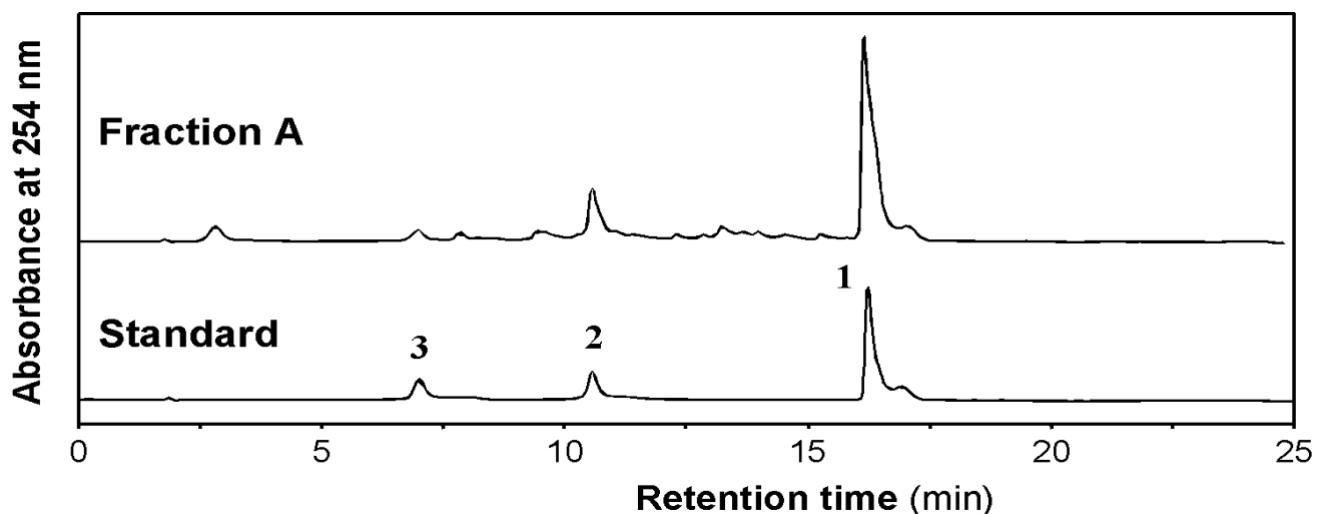


Figure 3. The constituents in Fraction A of moutan bark extract. HPLC chromatogram of Fraction A of moutan bark extract. Three constituents, paeonol (Compound **1**), β -PGG (**2**), and methyl gallate (**3**), were detected as described in the Materials and Methods. The number of each peak corresponds to the compound number.

Table 2. Constituents in moutan bark extract and their effects on NO production.

Constituent (compound)	Content in the fraction [%] ^a	Content in the extract [%] ^b	IC ₅₀ [μ M] ^c
MeOH extract	—	100	—
Fraction A (EtOAc-soluble fraction)	100	22.2	—
Paeonol (1)	32.5	7.22	208 \pm 77.3
1,2,3,4,6-Pentagalloyl- β -D-glucose (β -PGG; 2)	11.5	2.55	2.67 \pm 0.36
Methyl gallate (3)	1.47	0.326	100 \pm 35.8
Fraction B (<i>n</i> -butanol-soluble fraction)	100	9.93	—
Paeoniflorin (4)	14.9	1.48	NA

^aThe content of each constituent was evaluated by HPLC and is shown as the percentage of the dry weight of the relevant crude fraction.

^bThe content in the extract was calculated by multiplying the yield of each fraction. ^cThe half-maximal inhibitory concentration of NO production in IL-1 β -stimulated hepatocytes (mean \pm SD). At least three experiments were performed to determine these values. NA, not applicable due to low activity.

The constituents of moutan bark suppress NO production in hepatocytes: NO production was measured in hepatocytes to examine the anti-inflammatory activity of the constituents in moutan bark extract. IL-1 β induced NO production, whereas NO was not produced in the absence of IL-1 β (Figure 4A). The three hydrophobic constituents in moutan bark Fraction A suppressed NO production, and β -PGG was the most effective compound. In the absence of IL-1 β , no constituent alone increased NO production. In contrast, paeoniflorin showed a weak ability to suppress NO

production in hepatocytes (data not shown), and its IC₅₀ value could not be calculated. None of these constituents were toxic to hepatocytes because LDH activity was very low in the culture medium to which each constituent was added (data not shown). When compared with the effect of loxoprofen, which is a nonsteroidal anti-inflammatory drug and a positive control [14], β -PGG more efficiently suppressed NO production (Figure 4B). In contrast, paeoniflorin had a low ability to suppress NO production, and the IC₅₀ value could not be calculated (data not shown).

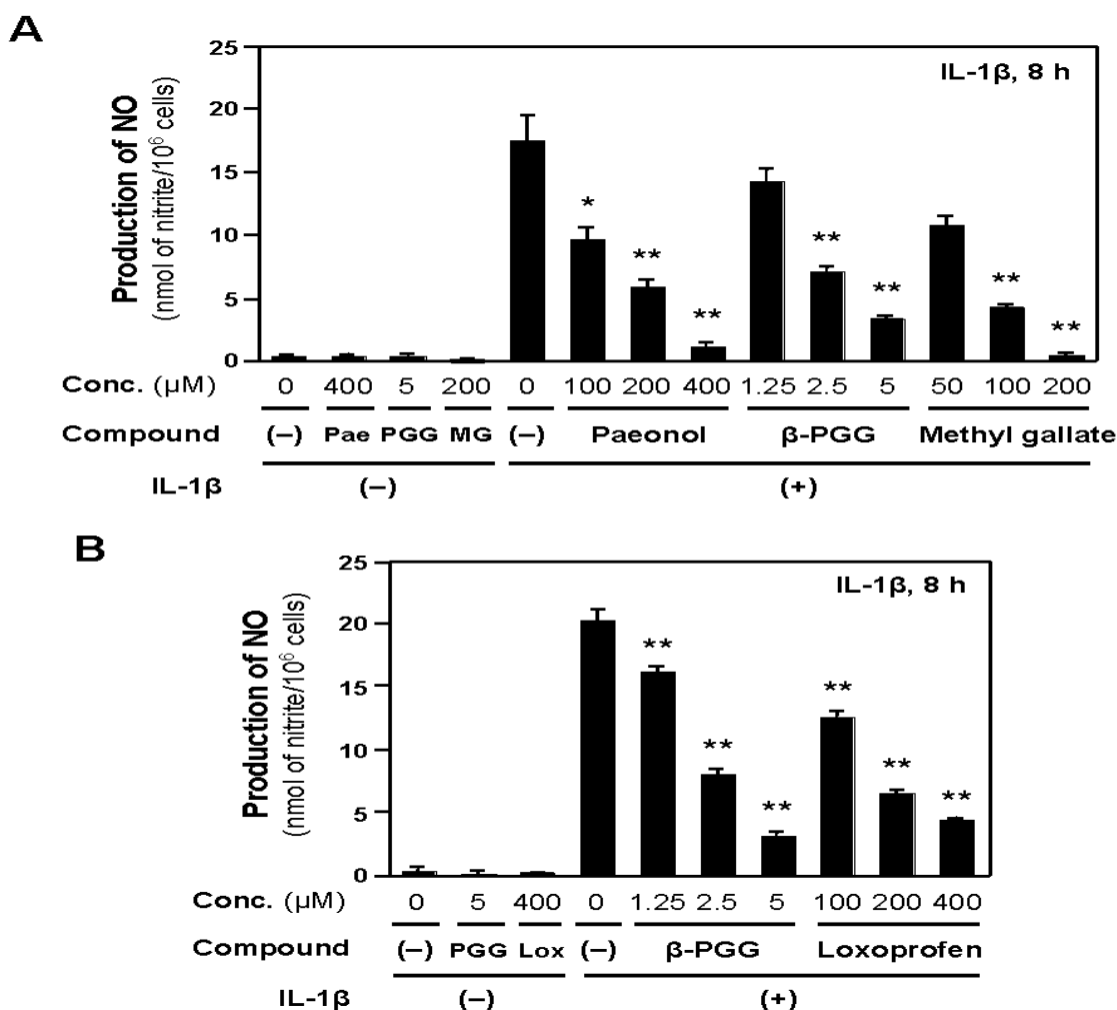


Figure 4. Hydrophobic constituents in moutan bark inhibit NO production in hepatocytes. (A) The effect of the constituents in Fraction A of moutan bark on NO production. Hepatocytes were treated with each constituent (paeonol, β -PGG, or methyl gallate (MG)) with/without IL-1 β for 8 h. Each constituent (Pae, PGG, and MG) was individually added to the medium as a negative control. The concentration of nitrite (a major metabolite of NO) in the medium was measured. The values are presented as the mean \pm SD ($n = 3$). * $P < 0.05$ and ** $P < 0.01$ versus IL-1 β alone. **(B)** The effect of β -PGG and loxoprofen on NO production. Loxoprofen (Lox) was added to the medium of hepatocytes as a positive control, and its ability to inhibit NO production was compared with that of β -PGG. Similar to (A), the NO concentration in the medium was measured and presented.

Next, the NO-quenching activity of moutan bark constituents was examined to rule out the possibility of direct NO quenching. The nitrite concentration was not significantly changed by any constituent (paeonol, β -PGG, or methyl gallate) compared with that in medium containing NaNO_2 alone (data not shown), suggesting that these constituents did not directly quench NO.

Hydrophobic constituents inhibit *iNOS* gene expression

in hepatocytes: We investigated whether moutan bark constituents downregulated *iNOS* gene expression in IL-1 β -stimulated hepatocytes. Among the constituents, β -PGG induced the most robust decrease in the protein expression of *iNOS* (Figure 5). Quantitative RT-PCR analysis showed that all constituents reduced *iNOS* mRNA levels in a concentration-dependent manner. In the absence of IL-1 β , *iNOS* mRNA levels were very low and could not be measured in hepatocytes (data not shown).

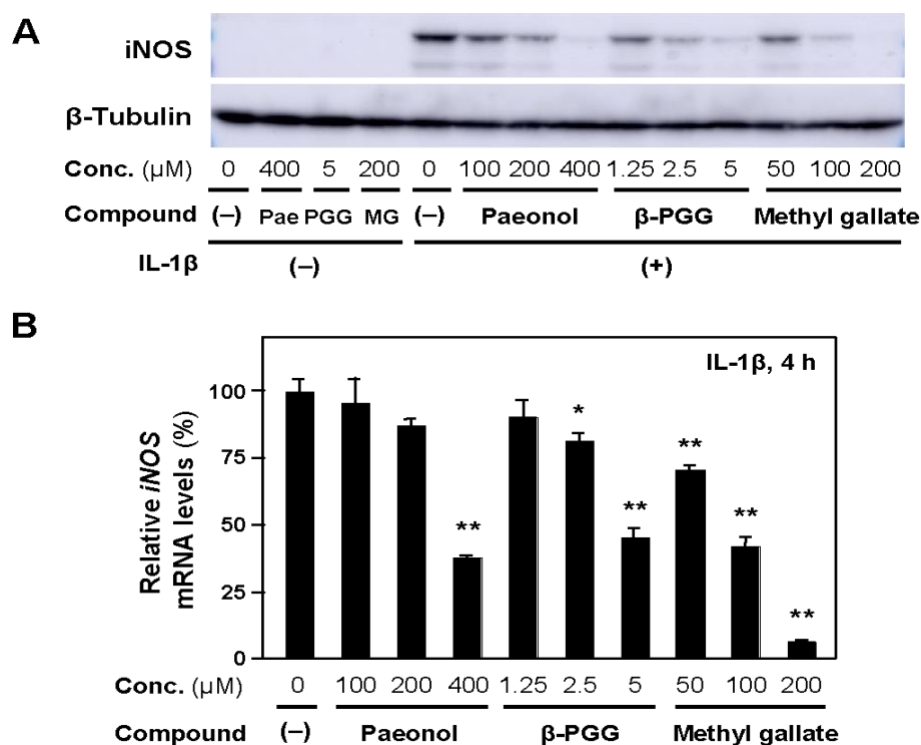


Figure 5. Hydrophobic constituents in moutan bark reduce *iNOS* gene expression. (A) The effect of moutan bark constituents on *iNOS* protein levels in hepatocytes. Cell extracts were prepared from the hepatocytes treated as described in Figure 4 (A) and subjected to Western blot analysis to visualize *iNOS* (130 kDa) and β -tubulin (55 kDa; internal control) protein expression. (B) The effects of moutan bark constituents on *iNOS* gene expression. Each constituent was added with IL-1 β and incubated for 4 h, and total RNA was analyzed by quantitative RT-PCR. The levels of *iNOS* mRNA were measured and normalized to *EF* mRNA (internal control). The normalized mRNA levels in the total RNA from hepatocytes incubated with IL-1 β alone were set as 100%. The relative mRNA levels are presented as the mean \pm SD ($n = 3$) as a percentage. * $P < 0.05$ and ** $P < 0.01$ versus IL-1 β alone.

Hydrophobic constituents induce anti-inflammatory effects on hepatocytes: Finally, we evaluated the inhibition of mRNA expression of genes involved in inflammation. Paeonol, β -PGG, and methyl gallate decreased the levels of TNF- α and IL1R1 mRNA in a concentration-dependent manner in IL-1 β -stimulated

hepatocytes (Figure 6). Furthermore, the hydrophobic constituents decreased the levels of IL-6, TNF- α , and chemokine C-C motif ligand (CCL) 20 mRNA (data not shown). These results suggest that these constituents, especially β -PGG, are responsible for the anti-inflammatory effects of moutan bark

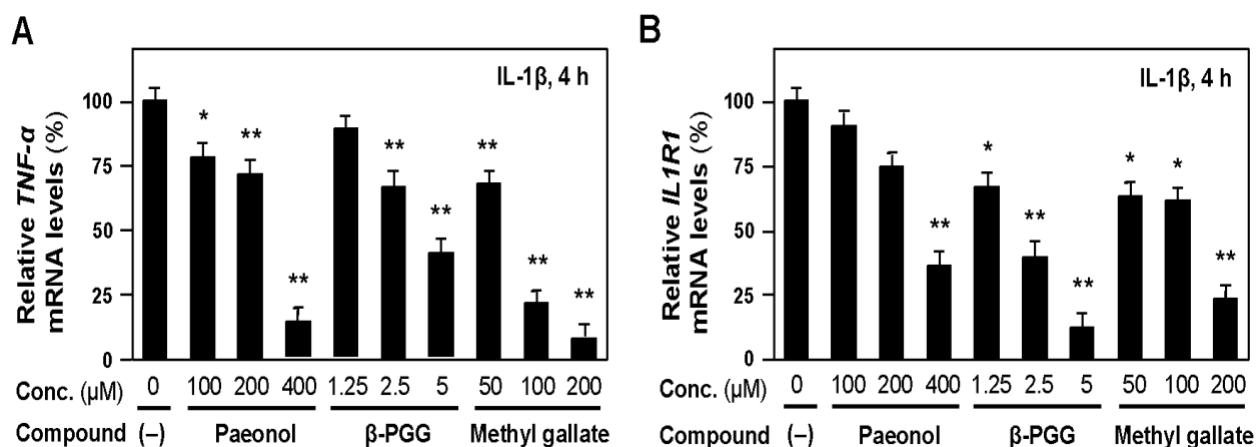


Figure 6. Moutan bark constituents decrease the mRNA expression of proinflammatory genes. (A) The effects of hydrophobic constituents of moutan bark on *TNF-α* mRNA expression in IL-1 β -treated hepatocytes. Total RNA was isolated from hepatocytes treated as shown in Figure 5 (B) and subjected to quantitative RT-PCR. The relative mRNA levels of *TNF-α* were measured and normalized to *EF* mRNA levels and are presented as the mean \pm SD ($n = 3$) in percent. * $P < 0.05$ and ** $P < 0.01$ versus IL-1 β alone. **(B)** The effects of moutan bark constituents on *IL1R1* mRNA expression. The relative mRNA levels of *IL1R1* were normalized to *EF* mRNA and are presented as the mean percent \pm SD ($n = 3$) in percent.

DISCUSSION

In this study, we identified the constituents that are characteristic of the root cortex of *Paeonia suffruticosa* (moutan bark) and determined the contents of these constituents using HPLC. As expected, paeonol and paeoniflorin were abundant in Fractions A and B, respectively (Table 1). Given that the yield of the MeOH extract of dried moutan bark was 25.9% (Figure 1) and the content in Fraction A of the extract was 22.2% (Table 1), the content of paeonol in moutan bark was calculated as 1.87% (18.7 mg/g of dried moutan bark). The paeonol content was comparable to previously reported results (13.08–14.29 mg/g dried moutan bark) [23]. The content of paeoniflorin was 14.9% of Fraction B, as determined by HPLC analysis, confirming that paeoniflorin was another major constituent.

An *ex vivo* hepatocyte system clearly demonstrated the potency of the constituents of moutan bark and that β -PGG markedly inhibited IL-1 β -induced gene expression of *iNOS* (Figure 5). According to the IC_{50} values, β -PGG (2.67 μ M) was 78-fold more potent than paeonol in

suppressing NO production (Table 1). The IC_{50} value of β -PGG was comparable to that of obakunone (2.6 μ M), which is in the bark of *Phellodendron amurense* [24]. Among the constituents of crude drugs that we examined using the *ex vivo* hepatocyte system, these IC_{50} values were the lowest.

β -PGG consists of five gallic acids and one glucose. Previously, we reported that the IC_{50} value for gallic acid was 212 μ M using the hepatocyte system [25], and the IC_{50} value for methyl gallate was 100 μ M (this study). Gallic acid is also known as a bioactive compound of a functional food [26]. In contrast, the IC_{50} value for glucose was not determined (not applied) due to low suppression activity [11]. Although further structure-activity relationships using galloyl glucose-derived compounds should be studied, the structure of gallic acid may contribute to the suppression of NO production in hepatocytes.

The signaling pathway initiated by IL1R1 is not well studied. In IL-1 β -stimulated human chondrocytes, paeonol exerted anti-inflammatory effects by inhibiting

the phosphatidyl inositol 3-kinase (PI3K)–Akt signaling pathway, leading to a decrease in NF- κ B activity [5, 27]. In LPS-treated macrophages, β -PGG decreased Toll-like receptor 4 (TLR4)-mediated activation of mitogen-activated protein kinase (MAPK) signaling pathway through NF- κ B by direct interaction with MyD88, which is an adaptor protein for both TLR4 and IL1R1 [28]. Although the hydrophobic constituents (paeonol, β -PGG, and methyl gallate) downregulated IL-1 β -induced expression of proinflammatory genes, the mechanism of this downregulation in hepatocytes should be elucidated in the future.

The inhibition of NO production is generally correlated with anti-inflammatory activities, such as the suppression of proinflammatory cytokine expression [9–10]. It has been reported that the IL-1 signal transduction pathway upregulates the expression of proinflammatory genes encoding iNOS, TNF- α , and IL-6 [17–18, 27]. Because the hydrophobic constituents of moutan bark decreased IL-1 β -induced expression of IL1R1 mRNA in hepatocytes (Figure 6), it is plausible that these factors inhibited the expression of genes downstream of IL1R1. Similar to our findings, β -PGG suppressed the secretion of inflammatory cytokines and chemokines, including CCL12 (also known as monocyte chemotactic protein 5) and pro-matrix metalloproteinase 9, in LPS-induced BV-2 cells, a mouse microglial cell line [29].

β -PGG, a simple hydrolysable tannin, is included in several crude drugs (*e.g.*, moutan bark), fruits, and nuts and thought to possess a variety of pharmacological activities, including anticancer and antidiabetic activities [30–31]. β -PGG binds to the insulin receptor and stimulates glucose uptake in mouse adipocyte-like 3T3-L1 cells [32]. Furthermore, PGG in the root of *Paeonia lactiflora* (the anomer of PGG, not indicated) functioned as an inhibitor of protein tyrosine phosphatase 1B (PTP1B), which negatively regulates insulin signaling [33]. Detailed studies on the pharmacological effects of the hydrophobic constituents of moutan bark are in progress.

CONCLUSION

This study indicated the hydrophobic constituents of the root cortex of *Paeonia suffruticosa* (β -PGG, methyl gallate, and paeonol) suppressed IL-1 β -induced expression of iNOS and proinflammatory genes. These constituents may be primarily responsible for the anti-inflammatory effects of this crude drug.

List of Abbreviations: NO: nitric oxide, IL: interleukin, TNF- α : tumor necrosis factor α , LPS: lipopolysaccharide, β -PGG: 1,2,3,4,6-pentagalloyl- β -D-glucose, iNOS: inducible nitric oxide synthase, IL1R1: IL-1 receptor, type 1, MeOH: methanol, EtOAc: ethyl acetate, i.d.: internal diameter, NMR: nuclear magnetic resonance, LDH: lactate dehydrogenase, EF: elongation factor 1 α .

Author contributions: MN and YI designed the study; TS, SS, and TO performed the experiments and analyzed the data; MN, YI, and SS prepared the manuscript; and all authors were involved in the performance of the experiments and preparation of the manuscript.

Competing Interests: TS performed the experiments as a graduate student of the Graduate School of Life Sciences, Ritsumeikan University. No authors have financial interests or conflicts of interest.

Acknowledgments and Funding: We thank Dr. Yuji Hasegawa (Daiichi University of Pharmacy) for mass spectral analyses; Ms. Aoi Shirai, Hiroko Uchida, and Airi Fujii for technical assistance; Springer Nature Author Services (<https://authorservices.springernature.com>) for editing a draft of this manuscript; and Ms. Noriko Kanazawa for her secretarial assistance. SS was supported by the Scholarship Fund for Young/Women Researchers of the Promotion and Mutual Aid Corporation for Private Schools of Japan. This work was partly supported by the Asia-Japan Research Institute, Ritsumeikan University.

REFERENCES

- The Committee on the Japanese Pharmacopoeia: *The Japanese Pharmacopoeia, 18th edn*. The Minister of Health, Labour and Welfare, Japan, 2021. <https://www.pmda.go.jp/english/rs-sb-std/standards-development/jp/0029.html>
- Wang Z, He C, Peng Y, Chen F, Xiao P: Origins, Phytochemistry, Pharmacology, Analytical Methods and Safety of Cortex Moutan (*Paeonia suffruticosa* Andrew): A Systematic Review. *Molecules* 2017, 22: 946. <https://doi.org/10.3390/molecules22060946>
- Chun SC, Jee SY, Lee SG, Park SJ, Lee JR, Kim SC: Anti-inflammatory activity of the methanol extract of moutan cortex in LPS-activated Raw264.7 cells. *Evid Based Complement Alternat Med* 2007, 4: 327–333. <https://doi.org/10.1093/ecam/nel093>
- Colasanti M, Suzuki H: The dual personality of NO. *Trends Pharmacol Sci* 2000, 21: 249–252. [https://doi.org/10.1016/S0165-6147\(00\)01499-1](https://doi.org/10.1016/S0165-6147(00)01499-1)
- Lou Y, Wang C, Tang Q, Zheng W, Feng Z, Yu X, Guo X, Wang J: Paeonol inhibits IL-1 β -induced inflammation via PI3K/Akt/ NF- κ B pathways: *In vivo* and *in vitro* studies. *Inflammation* 2017, 40, 1698–1706. <https://doi.org/10.1007/s10753-017-0611-8>
- Zhang L, Wei W: Anti-inflammatory and immunoregulatory effects of paeoniflorin and total glucosides of paeony. *Pharmacol Ther* 2020, 207: 107452. <https://www.doi.org/10.1016/j.pharmthera.2019.107452>
- Cao Y, Himmeldirk KB, Qian Y, Ren Y, Malki A, Chen X: Biological and biomedical functions of Penta-O-galloyl-D-glucose and its derivatives. *J Nat Med* 2014, 68: 465–472. <https://doi.org/10.1007/s11418-014-0823-2>
- Kitade H, Sakitani K, Inoue K, Masu Y, Kawada N, Hiramatsu Y, Kamiyama Y, Okumura T, Ito S: Interleukin 1 β markedly stimulates nitric oxide formation in the absence of other cytokines or lipopolysaccharide in primary cultured rat hepatocytes but not in Kupffer cells. *Hepatology* 1996, 23: 797–802. <https://doi.org/10.1053/jhep.1996.v23.pm0008666334>
- Ningsih FN, Okuyama T, To S, Nishidono Y, Okumura T, Tanaka K, Ikeya Y, Nishizawa M: Comparative analysis of anti-inflammatory activity of the constituents of the rhizome of *Cnidium officinale* using rat hepatocytes. *Biol Pharm Bull* 2020, 43: 1867–1875. <https://doi.org/10.1248/bpp.b20-00416>
- Yamauchi Y, Okuyama T, Ishii T, Okumura T, Ikeya Y, Nishizawa M: Sakuranetin downregulates inducible nitric oxide synthase expression by affecting interleukin-1 receptor and CCAAT/enhancer-binding protein β . *J Nat Med* 2019, 73: 353–368. <https://doi.org/10.1007/s11418-018-1267-x>
- Ohno N, Yoshigai E, Okuyama T, Yamamoto Y, Okumura T, Sato K, Ikeya Y, Nishizawa M: Chlorogenic acid from the Japanese herbal medicine Kinginka (*Flos Lonicerae japonicae*) suppresses the expression of inducible nitric oxide synthase in rat hepatocytes. *HOAJ Biology* 2012, 1: 2. <https://doi.org/10.7243/2050-0874-1-2>
- Kanemaki T, Kitade H, Hiramatsu Y, Kamiyama Y, Okumura T: Stimulation of glycogen degradation by prostaglandin E₂ in primary cultured rat hepatocytes. *Prostaglandins* 1993, 45: 459–474. [https://doi.org/10.1016/0090-6980\(93\)90122-N](https://doi.org/10.1016/0090-6980(93)90122-N)
- Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR: Analysis of nitrate, and [¹⁵N]nitrate in biological fluids. *Anal Biochem* 1982, 126: 131–138. [https://doi.org/10.1016/0003-2697\(82\)90118-X](https://doi.org/10.1016/0003-2697(82)90118-X)
- Inaba H, Yoshigai E, Okuyama T, Murakoshi M, Sugiyama K, Nishino H, Nishizawa M: Antipyretic analgesic drugs have different mechanisms for regulation of the expression of inducible nitric oxide synthase in hepatocytes and macrophages. *Nitric Oxide* 2015, 44: 61–70. <https://doi.org/10.1016/j.niox.2014.12.001>
- Tanemoto R, Okuyama T, Matsuo H, Okumura T, Ikeya Y, Nishizawa M: The constituents of licorice (*Glycyrrhiza uralensis*) differentially suppress nitric oxide production in interleukin-1 β -treated hepatocytes. *Biochem Biophys Res* 2015, 2: 153–159. <https://doi.org/10.1016/j.bbrep.2015.06.004>
- Takimoto Y, Qian HY, Yoshigai E, Okumura T, Ikeya Y, Nishizawa M: Gomisin N in the herbal drug gomishi (*Schisandra chinensis*) suppresses inducible nitric oxide synthase gene via C/EBP β and NF- κ B in rat hepatocytes. *Nitric Oxide* 2013, 28: 47–56. <https://doi.org/10.1016/j.niox.2012.10.003>
- Matsui K, Nishizawa M, Ozaki T, Kimura T, Hashimoto I, Yamada M, Kaibori M, Kamiyama Y, Ito S, Okumura T: Natural antisense transcript stabilizes inducible nitric oxide synthase messenger RNA in rat hepatocytes. *Hepatology* 2008, 47: 686–697. <https://doi.org/10.1002/hep.22036>
- Yoshigai E, Hara T, Inaba H, Hashimoto I, Tanaka Y, Kaibori M, Kimura T, Okumura T, Kwon AH, Nishizawa M: Interleukin-1 β induces tumor necrosis factor- α secretion from rat hepatocytes. *Hepatol Res* 2014, 44: 571–583.

- <https://doi.org/10.1111/hepr.12157>
19. Ding L, Liu Z, Zhao F, Bai G, Chen L, Yao X, Qiu F: Isolation and identification of the metabolites of paeonol in human urine. *Xenobiotica* 2012, 42, 1206–1212. <https://doi.org/10.3109/00498254.2012.694492>
 20. Mitsuhashi H, Hayashi K, Nomura T: Studies on the Constituents of Asclepiadaceae Plants. XVIII. Components of *Cynanchum paniculatum* KITAGAWA. *Chem Pharm Bull* 1966, 14: 779–783. <https://doi.org/10.1248/cpb.14.779>
 21. Zhang Y, DeWitt DL, Murugesan S, Nair MG: Novel lipid-peroxidation- and cyclooxygenase-inhibitory tannins from *Picrorhiza kurroa* seeds. *Chem Biodivers* 2004, 1: 426–441. <https://doi.org/10.1002/cbdv.200490036>
 22. Cheng KW, Yang RY, Tsou SCS, Lo CSC, Ho CT, Lee TC, Wang M: Analysis of antioxidant activity and antioxidant constituents of Chinese toon. *Journal of Functional Foods* 2009, 1: 253–259. <https://doi.org/10.1016/j.jff.2009.01.013>
 23. Yang G, Zhang C, Li P, Qiu Y, Dong F: Chemical fingerprinting and quantitative analysis of cortex moutan from different tree peony cultivars using HPLC-ESI/MS. *Natural Product Communications* 2020, 15: 11. <https://doi.org/10.1177/1934578X20973519>
 24. Fujii A, Okuyama T, Wakame K, Okumura T, Ikeya Y, Nishizawa M: Identification of anti-inflammatory constituents in Phellodendri Cortex and Coptidis Rhizoma by monitoring the suppression of nitric oxide production. *J Nat Med* 2017, 71: 745–756. <https://doi.org/10.1007/s11418-017-1107-4>
 25. Yamanishi R, Yoshigai E, Okuyama T, Mori M, Hiromitsu Murase H, Machida T, Okumura T, Nishizawa M: The Anti-Inflammatory Effects of Flavanol-Rich Lychee Fruit Extract in Rat Hepatocytes. *PLoS One* 2014, 9: e93818. <https://doi.org/10.1371/journal.pone.0093818>
 26. Martirosyan D, Lampert T, Morgan Ekblad M: Classification and regulation of functional food proposed by the Functional Food Center. *Functional Food Science* 2022, 2: 25–46. <https://doi.org/10.31989/ffs.v2i2.890>
 27. Karin M: NF-κB as a critical link between inflammation and cancer. *Cold Spring Harb Perspect Biol* 2009, 1: a000141. <https://doi.org/10.1101/cshperspect.a000141>
 28. Jang SE, Hyam SR, Jeong JJ, Han MJ, Kim DH: Penta-O-galloyl-β-D-glucose ameliorates inflammation by inhibiting MyD88/NF-κB and MyD88/MAPK signalling pathways. *Br J Pharmacol* 2013, 170: 1078–1091. <https://doi.org/10.1111/bph.12333>
 29. Mendonca P, Taka E, Bauer D, Cobourne-Duval M, Soliman KFA: The attenuating effects of 1,2,3,4,6 penta-O-galloyl-β-D-glucose on inflammatory cytokines release from activated BV-2 microglial cells. *J Neuroimmunol* 2017, 305: 9–15. <https://www.doi.org/10.1016/j.jneuroim.2017.01.011>
 30. Smeriglio A, Barreca D, Bellocco E, Trombetta D: Proanthocyanidins and hydrolysable tannins: occurrence, dietary intake and pharmacological effects. *Br J Pharmacol* 2017, 174: 1244–1262. <https://doi.org/10.1111/bph.13630>
 31. Zhang J, Li L, Kim SH, Hagerman AE, Lü J: Anti-cancer, anti-diabetic and other pharmacologic and biological activities of penta-galloyl-glucose. *Pharm Res* 2009, 26: 2066–2080. <https://doi.org/10.1007/s11095-009-9932-0>
 32. Li Y, Kim J, Li J, Liu F, Liu X, Himmeldirk K, Yulin Ren Y, Wagner TE, Chen X: Natural anti-diabetic compound 1,2,3,4,6-penta-O-galloyl-D-glucopyranose binds to insulin receptor and activates insulin-mediated glucose transport signaling pathway. *Biochem Biophys Res Commun* 2005, 336, 430–437. <https://doi.org/10.1016/j.bbrc.2005.08.103>
 33. Baumgartner RR, Steinmann D, Heiss EH, Atanasov AG, Ganzera M, Stuppner H, Dirsch VM: Bioactivity-Guided Isolation of 1,2,3,4,6-Penta-O-galloyl-D-glucopyranose from *Paeonia lactiflora* Roots As a PTP1B Inhibitor. *J Nat Prod* 2010, 73: 1578–1581. <https://www.doi.org/10.1021/np100258e>