



Changes of volatile flavored compounds during lactic acid fermentation in cereal-based products

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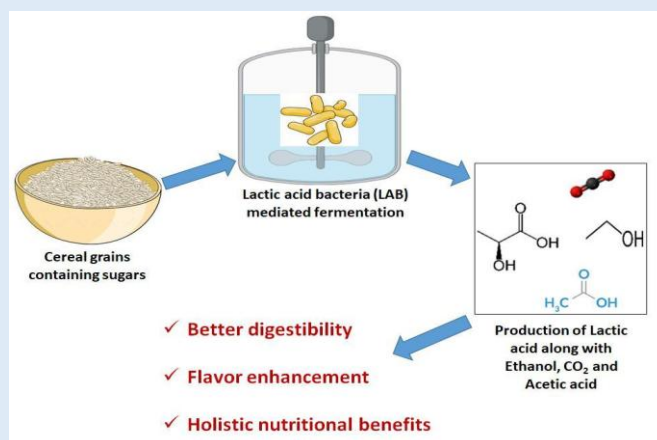
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ABSTRACT

Lactic acid bacteria (LAB)-mediated fermentation of cereals is a preservation technique that enhances food flavor, nutrition, and safety. LAB naturally exists in cereals and plays a significant role in fermentation converting sugars into lactic acid and lowering the pH levels to create a distinctive sour taste and aroma. This process increases nutritional value and produces compounds like ethanol, acetic acid, and carbon dioxide, contributing to texture and appearance. LAB fermentation breaks down complex carbohydrates and proteins, improving digestibility and nutrient absorption. Common LAB-fermented cereal-based foods include sourdough bread, fermented cereal porridges, yogurt and kefir. Effective monitoring of volatile flavor compounds during fermentation is crucial for ensuring the quality, safety, and consumer acceptance of these foods.

Keywords: Cereal-based food, Volatile flavor compounds, Lactic acid bacteria, Fermentation, Amino acid metabolism, HPLC, GC-MS



Graphical Abstract: Changes of volatile flavored compounds during lactic acid fermentation in cereal-based products

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INTRODUCTION

Consuming cereals has become a primary energy source providing essential protein, carbohydrates, fiber and minerals [1]. However, the application of cereal fermentation is limited due to the poor sensory properties. Three methods are employed to enhance nutritional quality: germination, milling, and fermentation. Among this fermentation, utilizing lactic acid bacteria (LAB) or probiotics, is the oldest and most effective. LAB fermentation increases cereal-based products' bioavailability, digestibility, and organoleptic properties [2]. During lactic acid fermentation (LAF), volatile flavor compounds undergo significant changes, impacting the final product's sensory appeal, appetite stimulations, and profile. The specific changes depend on the LAB strain and cereal used. LAB produces enzymes breaking down complex carbohydrates and proteins into smaller molecules, such as organic acids, (propionic acid, acetic and lactic acid) and metabolites (diacetyl, acetaldehyde, and ethanol). This study may provide insight into the changes in volatile flavor compounds during LAF in cereal-based products.

Process for Identification of Volatile Compounds: For identification of the volatile flavoring compounding compounds, the first step is fermentation. Modifications in the fermentation process can be made to yield diverse results. For instance, fermenting various cereals with the same microorganism [3-5] and or using the same cereal by different bacteriological bacterial or fungal strains on the same cereal [6-8] produces a range of similar volatiles with variations. Analyzing these variations and their concentrations help determine the optimal cereal-strain combination for maximum biotechnological benefits and pharmaceutical applications.

Mass spectrometry is employed to identify volatile compounds in fermented food products. The analytical method involves: Solid Phase Extraction (SPE), Gas

chromatography with mass spectrometry (GC-MS). The procedure includes the sorption of volatile compounds using 6-*amyl-a*-pyrone, at 900 rpm for one hour. The absorbed volatiles are then desorbed using a twister and thermal unit, followed by GC-MS analysis. By comparing the retrieved data with standard values [9-11] and retention indices, the evolved volatile compounds during fermentation can be identified.

Post-extraction derivatization and analysis of fatty acids in fermented cereals: Following extraction, fatty acids present in fermented cereal samples undergo derivatization. The sample is treated with heneicosanoic acid and vortexed with a 10:5:2 (v/v) methanol: chloroform: distilled water mixture, followed by sonication [12]. Subsequent steps include ultralow-temperature centrifugation, supernatant evaporation and concentration, and high-performance liquid chromatology (HPLC), followed by Gas chromatography-mass spectrometry (GC-MS) for structural confirmation.

Sensory evaluation of volatile compounds: Sensory evaluation is crucial for researchers studying volatile flavor compounds. Figure 1 represents the process for the identification of volatile organic compounds. Researchers use two tests, conducted by trained, unbiased judges, first to assign attributes based on the aftertaste, aroma, and original taste. The second test assigns a characteristic to each component, aiding in the organoleptic assessment of the flavor profile and identifying volatile compounds that impart flavor. This process identifies volatile compounds released during cereal fermentation, providing descriptive and quantitative data. The panel is informed of the range maxima and minima [13]—table 1 lists common characteristics assigned to attributes and is suitable for evaluating volatiles released upon cereal fermentation.

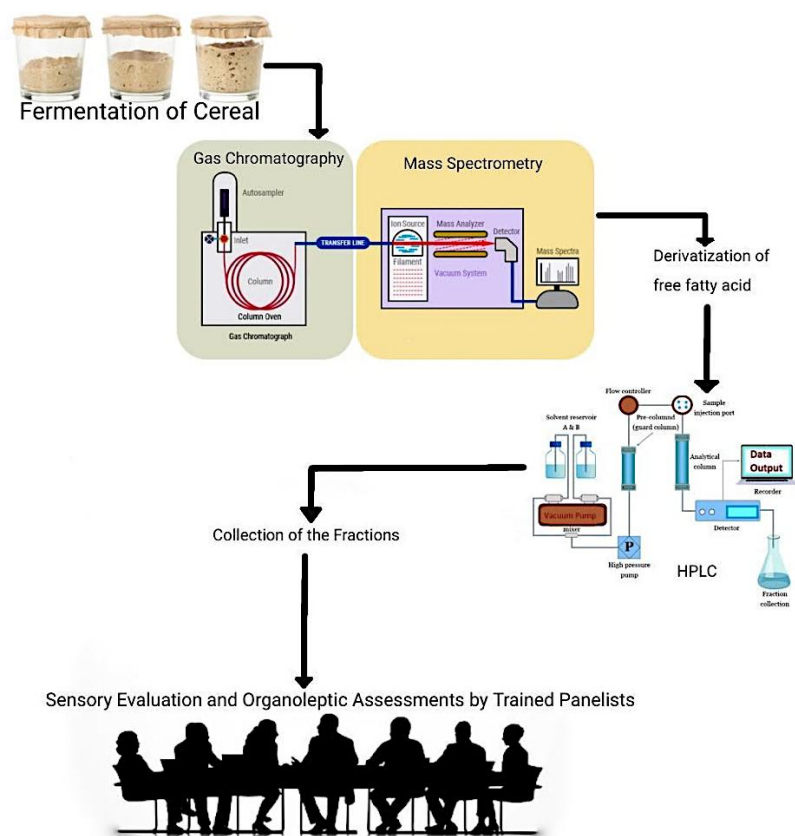


Figure 1: Process for identification of Volatile Organic Compounds

Table 1. Attribute based Characteristics assigned [14]

Characteristic(s) Assigned	Attribute	References
Sticky toffee like	Caramel	[13-14]
Grassy herb like, root like	Worty	
Foul-Drain	Sulphur	
Peach like, subtle	Fruity	
Floral	Apple like	
Taste sensation related to iron nails	Metallic	
Taste sensation related with sugary items	Sweet	
Caffeine/ tonic like	Bitter	
Warming, spirit like	Alcohol	
Malty/ raw Horlicks	Malt	
Citrusy, Lemony	Sour	
Stinging, Burning, Heat	Spicy	
Butterscotch like	Diacetyl	

Types of VCs (Volatile Compounds): Fermentation of cereal-based food items produces volatile flavoring compounds, with aldehyde and ketone-based compounds making up 70% of total volatile compounds. Other prominent compounds include benzene, hydrocarbons, furans, nitrogen, sulfur-containing compounds, and lactones. Aldehyde and ketones, derived from cereal lipids, exhibit tautomeric properties. Their concentrations vary among cereals, with brown rice

exhibiting higher levels due to its bran layer, while other cereals have a higher ratio of aldehydes and ketones attributed to their increased fat, lipid content, and presence of bran layers [15]. Studies show that lactones have a coconut-like, floral-fruity smell that intensifies during fermentation. [16]. Extensive literature review reveals that volatile compounds can be classified into 10 distinct segments/types. Table 2 summarizes the most reported volatile compounds within each type.

Table 2. Commonly reported volatile compounds

Type of Volatile	Compounds	References
Aldehyde	Methyl-1-butanal, hexanal, heptanal, (E)-2-hexanal, fural, benzaldehyde, octanal, (E,E)-2,4-heptandial	[3,4,6,7,8,14,17,18]
Ketones	Diacetyl, acetoin, Acetol, acetophenone, 6-methyl-3,5-heptadiene-2-one 2-heptone	
Alcohols	Ethanol, 2-methyl butanol, 2-methyl-1-propanol, hexanol, 2-butoxyethanol, octan-3-ol Benzomethanol	
Esters	Ethyl hexdecanoate, methyl hexdecanoate, Ethyl hexanoate, 1-methyl-2-hydroxypropanoate	
Hydrocarbons	Styrene, 1,3-xylene Pentadecane, heptadecane 1,4-Xylene, toluene, Lycopene	
Furans	2-Pentylfuran, Butylfuran, 2-propylfuran	
Phenols	(4-vinyl)-2meth-oxyphenols	
Sulphur rich	Benzothiazole	
Nitrogen rich	N,N-dibutylformldehyde	
Lactones	δ-dodecalactone, γ-decalactone, γ-nonolactone	

Pathway of formation of volatile compounds and effects of multiple strains on them: The fermentation process involves multiple metabolic pathways, including carbohydrate metabolism, which yields volatile compounds. The Embden-Meyerhof-Parnas pathway produces fermentation compounds like butanediol and ethanol, with upregulation of enzymes like dehydrogenase and decarboxylase observed via Heat Maps [19]. When compared to the microorganisms used for fermentation, reports state that concentrations of

aldehyde were greater when *Aspergillus oryzae* was used, while the use of *L. plantarum* increased the final quantities of acetic acid; however, none of these compounds were detected in the unfermented samples. An overall conclusion can be drawn that fermentation is carried out using fungi such as *S. cerevisiae*, *Ryzoopus* sp. *Aspergillus* sp. increased the total by-products that can be formed in the fermentation process of carbohydrates, particularly ethanol and ethyl acetate, rocketing at about 200 and 25 folds compared to the control.

When delved into the fermentation of butanediol, the volatiles acetoin i.e., 3-hydroxybutan-2-one and diacetyl - butane-2,3-dione needed the enzyme alpha-acetolactate decarboxylase and at the end the dehydration by the enzyme 2,3-butanediol dehydrogenase [20]. Literature portrays that higher yield of acetoin can be obtained by using various species of LAB [21]. Molecules such as sulphur and nitrogen can only be a part of the volatile by-products of fermentation if the metabolic pathway of utilizing amino acids is involved [22]. Nitrogen is sourced from amino acids, while sulfur comes from cysteine and methionine. Amino acids can produce branched chain alkanes upon metabolism, while aromatic rings can be found in tryptophan, phenylalanine, and tyrosine. Methylbutanal derivatives are detected more frequently when fungi are used than LAB, possibly due to a lack of weak vital enzymes [23].

Lipid metabolism is also a pathway that produces several VCs, such as lactones, furans, and many more, via

the degradation of fatty acids, ketone glycerol, and esters. The core conversion of these long carbon chain compounds into two carbon segments of acetyl-Co-A happens via beta-oxidation. At the same time, the following stages are carried out by specific enzymes [24]. In the sample that is fermented, the presence of unsaturated fat such as linolenic acid and oleic acids leads to increased Lactone formation using the β -oxidations and hydroxylation of the fatty acids [16], which imparts the products arising from these fermentations a fruity-floral odor. The fermented cereal-based products often have a strong smell that mimics the sweet scent of rose flowers. This is a demarcation feature of 2-phenylethanol [25]. Other volatile benzoic compounds, such as styrene, phenyl-ethan-1-one, are derived from phenylalanine-containing cinnamic acids [26]. Notably, 4-ethyl-2-methoxyphenol is the primary phenolic compound cited in the fermentation of rice and cereals, imparting a characteristic smoky odor. Different possible metabolic fates of biomacromolecules like sugars and amino acids are represented in Figure 2.

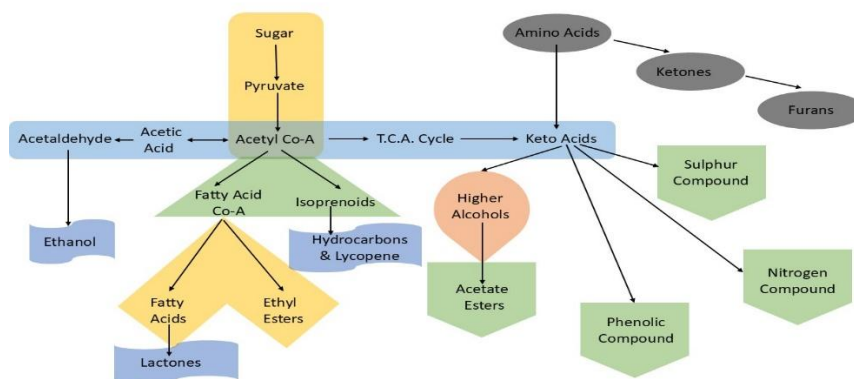


Figure 2. Metabolism of sugar and amino acids to pre-volatile compounds.

Various strains of LAB, along with its specific volatile compounds and its properties: Cereals, grown on 73% of global land, contribute over 60% of worldwide production providing essential nutrients, fiber, protein, energy, minerals, and vitamins. Fermented cereals, particularly in African and Asian countries, are integral to traditional foods like sourdough bread, injera, and ogi, offering highly nutritional value, cultural significance, and importance in global diets [18]. The cereal fermentation

process yields end and intermediate products with volatile aromatic compounds, contributing to a harmonious aroma and flavor. Key aroma compounds exhibit a dilution factor of flavor ≥ 16 , indicating their potency and significance in the overall appeal, taste, and smell of food. These compounds confer functional benefits to consumers. Functional foods are natural or processed items containing known or unknown biologically active compounds, providing clinically proven

health benefits for chronic disease prevention, management, or treatment. Key volatile compounds identified include vanillin, sotolon, 2-phenyl ethanol, β -damascenone, acetic acid, and furaneol [27,28]. Analytical methods involve microextraction with solid-phase in headspace, gas chromatography, and GC-MS

confirmation [3]. Alternatively, automatic headspace gas chromatography (HS-GC) is used, varying by cereal type [7]. Table 3 presents various lactic acid bacteria strains, and their corresponding volatile compounds produced during cereal fermentation.

Table 3: Strain of Bacteria and their Volatiles.

Strains of Bacteria	Cereals/Products	Volatile Compounds	Properties	Reference
<i>Lactobacillus bulgaris</i>	Rice, maize	Ester, ethanols	Aromatic compounds	[8]
<i>Streptococcus thermophilus</i>	Rice, Maize	Ester, ethanols	Aromatic compounds	[8]
<i>Lactobacillus plantarum</i>	Maize, millet sorghum Cocoa, Rice	(Aldehydes, ketones, esters and alcohols	Aroma and flavors	[29]
<i>L. paracasei</i>	Rice, Millet	γ -decalactone, γ -nonalactone, β -lactones, δ -lactones	Flavor and Aroma	[16]
<i>Enterococcus genera.</i>	Sorghum, finger millet	Ethanol, Lactic acid	Alcoholic flavored contribution to bushera	[6,7]
<i>Lactobacillus paracasei</i>	maize	2,3-butanedione and 2-butanone, 3-hydroxy	Flavor compounds	[4]
<i>Lactobacillus fermentum</i>	Maize millet sorghum Cocoa	(Aldehydes, ketones, esters and alcohols	Flavor and aroma	[4,29]
<i>Lactobacillus brevis</i>	Togwa production	3-methyl-1-butan-1-ol, 2-methyl-1-propan-1-al, 3-methyl-1-butanal, 2-methyl-(1)-butanal, 2-methyl butan-1-ol and 2-methyl-1-propan-1-ol		[3]
<i>Lactobacillus delbrueckii subsp. bulgaricus</i>	Maize, millet, kefir grains	hydrocarbons, ketone, ester, aldehydes, organic acid, alcohols, compounds containing sulfur and benzoic compounds	Flavor compounds,	[5]
<i>Lactobacillus spp.</i>	White rice, brown rice	1-octanol 1-hexanol, 1-heptanol, nonanol	Aromas and Flavor	[17]
<i>Lactococcus lactis</i>	Quinoa, oats, barley, rice	Ketone, aldehyde	Aromas and flavor	[4]
<i>Lactobacillus cellobiosus</i>	Togwa	3-methyl-1-butan-1-ol, 2-methyl-1-propan-1-al, 3-methyl-1-butanal, 2-methyl-(1)-butanal, 2-methyl butan-1-ol and 2-methyl-1-propan-1-ol	Helps to yield butanoic acid	[3]
<i>Pediococcus pentosaceus</i>	Emmer	3-methyl-1-butan-1-ol, 2-methyl-1-propan-1-al, 3-methyl-1-butanal, 2-methyl-(1)-butanal, 2-methyl butan-1-ol and 2-methyl-1-propan-1-ol	Helps to yield butanoic acid	[3]
<i>Lactobacillus casei</i>	Sorghum, rice, glutinous rice, wheat bran	Acetoin	Has a key role in formation of the dominant flavor in cereal vinegars	[18]
<i>Acetobacter pasteurianus</i>	Sorghum, rice, glutinous rice, wheat bran	Acetoin	It has a crucial aspect in forming the dominant flavor in cereal vinegars	[18]

Strain-specific contribution to the fermentation of cereals: Fermentation with LAB is a centuries-old preservation process that converts sugars into lactic acid, lowering the food pH and creating a hostile environment for spoilage and pathogenic microorganisms. This process extends shelf life and enhances the nutritional value and sensory properties of fermented foods. LAB produces antimicrobial metabolites like acetic acid, propionic acid, and bacteriocins, *Lactiplantibacillus plantarum* and *Lactocaseibacillus rhamnosus* are conditionally heterofermentative, while *Lactocaseibacillus casei* and *Pediococcus pentosaceus* are predominantly homofermentative, and *Levilactobacillus brevis* is heterofermentative [30].

In cocoa fermentation, a complex microbiota of acetic acid bacteria, yeasts, and LAB produces key components like acetic acid, lactic acid, and ethanol, contributing to flavor and aroma. Secondary metabolites, such as esters, alcohols, and acids, shape the complex flavor profile, dependent on strain, temperature, pH, and pathway. These metabolites are crucial for developing chocolate's potent aroma and characteristic flavor. The diversity of microbial populations and fermentation conditions influence the sensory profile. Furthermore, enzymes responsible for malt and fermenting microorganisms exploit their proteolytic activity to create precursors of aromatic compounds, such as amino acids. Optimal fermentation microorganisms have been identified. For instance, *Streptococcus thermophilus* Conversely, *Pediococcus spp.* and *Lactobacillus plantarum* dominate corn dough fermentation, causing rapid acidification [18].

Flour from corn, oats, and barley has been extensively reported in literature for producing

fermentable cereal-based products. LAB play a crucial role in developing desirable properties in cereal- and pseudocereal-based beverages, including lipid conversion, water-soluble vitamin synthesis (B vitamins), enzymatic breakdown of phytic acid, and enhanced protein digestibility. Pseudocereal-based drinks can also be fortified through mechanisms beyond phytic acid polyphenol degradation, with LAB fermentation altering protein and amino acid content [4]. The food industry is developing functional beverages to meet growing demand for health-benefiting and disease-reducing products. Fermented foods, alcoholic and non-alcoholic beverages based on sorghum or millet, are gaining popularity. For instance, Obutoko type obushera (sorghum koji) made from Secedo red sorghum (*Sorghum bicolor* (*L. Moench*)) flour exhibits signature ethanol production and bold alcoholic taste [7].

Impacts of using Volatile compounds: Naturally present in fermented products, volatile compounds offer preservative properties [31], potentially reducing chemical preservative use. These compounds have been linked to cancer prevention [32], type 2 diabetes management [33-34], and neurodegenerative disorder prevention [35], potentially reducing the use of chemical-based preservatives. Antimicrobial activity, seen in (E)-2-hexen-1-al, makes them valuable biocontrol agents against pathogens [36]. However, some VOCs may pose health risks, including ENT irritation, headaches, nausea, kidney damage, liver damage, cancer, cardiac arrhythmia, vagal inhibition, respiratory system collapse, and sudden death [37]. Fortunately, their production in fermentation processes is typically negligible and statistically insignificant

Table 4: VOC containing Functional foods in health

Food	Cereal	Fermentation Process	Health Benefits
Idli and Dosa	Rice, Urad Dal	Lactic acid fermentation	Rich in probiotics, aids digestion reduce obesity [35]
Tempeh	Soybeans	Fungal fermentation	High in protein, fiber, and vitamins, especially B vitamins [35]
Miso	Soybeans, rice, barley	Fungal fermentation	High in protein, fiber, and antioxidants, supports gut health
Natto	Soybeans	Bacterial fermentation	High in protein, fiber, and vitamin K2, aids bone health [34]
Kefir	wheat, barley	Bacterial and yeast fermentation	Probiotic-rich, supports gut health, improves digestion [36]
Sourdough Bread	Wheat, rye,	Lactic acid and yeast fermentation	Easier to digest, may improve gut health, contains beneficial prebiotics [35]
Beer	Barley	Yeast fermentation	Contains antioxidants and polyphenols, reduces risk of heart disease and certain cancers [38]
Wine	Grapes	Yeast fermentation	Contains antioxidants and polyphenols [4]

Fermentation of cereals and legumes alters antioxidant levels and properties, transforming them into functional food ingredients. Research has consistently shown that this process imparts anti-obesity properties [4,35,38-39]. Moreover, the probiotics and prebiotics resulting from lactic acid fermentation (LAF) have been found to alleviate hangover symptoms and support live health, preventing diseases and disorders [32-34,40]. Black Mahalab, a cereal seed native to subtropical and tropical regions, has been a staple in Sudanese traditional meals for centuries. Fermentation is a key cooking method, rendering it a functional food component. Studies have revealed its antimicrobial and laxative properties, relieving irritable bowel syndrome (IBS) and other stomach conditions [17,41]. These examples demonstrate the potential of fermented cereal-based foods as functional ingredients, highlighting their role in maintaining health and combating disease. These foods have contributed to human well-being for centuries, underscoring the importance of preserving traditional fermentation practices and exploring their modern applications.

Functional Effects of Fermented Cereal VOCs- Prospects of Use: Research volatile flavorings have revealed promising potential in disease prevention, yet the

industry has been slow to develop these compounds for therapeutic applications [42-43].

However, growing interest in antimicrobials found in fermented cereal-based foods has sparked innovation in functional food development, positioning this area as a hub for health-focused research [35,44]. Fermented plant-based foods are an abundant source of volatile aromatic flavoring compounds, which have shown impressive results in improving shelf life [45], enhancing health benefits [46,47], exhibiting fungicidal and insecticidal properties domain [43,48,49], and defending against microorganism-based decay in wounds (animals and plants) [45].

The antimicrobial effects of these volatiles are attributed to the simultaneous interactions of multiple volatile molecules, rather than single compounds [46]. Three major categories of volatiles with preservative potential have been identified: phenolic and phenol compounds, alkaloids, and terpenoids [50]. These findings underscore the vast potential of volatile compounds in fermented foods, warranting further research and development in the food-pharma interface.

Research has unveiled the antimicrobial potential of various volatile compounds found in foods. For instance, E-2-hexenal, an aldehyde from raw green cocoa beans,

exhibits potent antimicrobial activity against pathogens [51]. Linalol and α -Terpineol, volatiles from barley and hop, inhibit the growth of cariogenic microorganisms and periodontopathic bacterial species [38]. Linalol's anti-histaminic and anti-inflammatory properties make it a valuable ingredient in pharmaceuticals [52] while its precursor, non-volatile geraniol, has been proposed as a component for treating neurological disorders [47,53]. Fermented beans and cereals contain significant bioactive volatiles, including vanillin, vanillic acid, and alcohol derivatives. These compounds are renowned for their anticarcinogenic properties, anticlastogenic properties [54], respiratory inhibition of pathogenic species, such as *L. plantarum*, *L. innocua*, and *E. coli* [55], and promising inhibitory effect on sickle cell disease. The pharmaceutical industry utilizes these compounds as natural flavoring agents and bioactive ingredients, harnessing their potential to develop innovative treatments.

CONCLUSION

Whole wheat and wheat-based cereals offer significant nutritional value, boasting dietary antioxidants and high esterified phenolic acid content. Fermented barley, rich in β -glucan, has been shown to control hypoglycemia and hypercholesterolemia, while also reducing colon cancer risk associated with harmful food chemicals. Other cereals, such as sorghum, finger millet, and oats, serve as excellent prebiotics, providing long-term bacterial support. India is home to various fermented functional foods, including idli, dosa, dhokla, selroti, khaman, ambeli, sez, vada, anarshe, balam, kurdi, naan, bhatura, kulcha, and jalebi. These fermented pulses and cereals enhance total protein content, oil-holding capacity, sensory attributes, volatile saponins, and phenolics, all essential for overall health support. LAF, a process where bacteria convert sugars into lactic acid, is integral to fermented foods like yogurt, sauerkraut, and kimchi. This

process yields volatile flavoring compounds, including aldehydes, alcohols, ketones, benzoids, acids, and sulphur. The type and amount of volatile organic compounds (VOCs) produced depend on the bacteria involved and cereal-derived food type.

Abbreviations: LAB, Lactic acid bacteria; LAF, lactic Acid fermentation; GC-MS, Gas chromatography/Mass spectrometry; Rpm, Revolutions per minute; HPLC, High Performance Liquid Chromatography; VOCs, Volatile organic compounds.

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REFERENCES

1. Ashagrie H, Baye K, Guibert B, Seyoum Y, Rochette I, Humblot C. Cereal-based fermented foods as a source of folate and cobalamin: The role of endogenous microbiota. *Food Res Int.* 2023 Dec;174(Pt 1):113625. DOI: <https://doi.org/10.1016/j.foodres.2023.113625>
2. Enujiugha, V. N., Badejo, A. A. Probiotic potentials of cereal-based beverages. *Critical Reviews in Food Science and Nutrition* 2017; 57(4), 790–804. DOI: <https://doi.org/10.1080/10408398.2014.930018>
3. Mugula, J. K., Narvhus, J. A., Sørhaug, T. Use of starter cultures of lactic acid bacteria and yeasts in the preparation of togwa, a Tanzanian fermented food. *International journal of food microbiology* 2003, 83(3), 307-318 DOI: [https://doi.org/10.1016/s0168-1605\(02\)00386-0](https://doi.org/10.1016/s0168-1605(02)00386-0)
4. Ziarno, M., Cichońska, P. Lactic acid bacteria-fermentable cereal-and pseudocereal-based beverages. *Microorganisms* 2021, 9(12), 2 DOI: <https://doi.org/10.3390/microorganisms9122532>

5. Yépez, A., Russo, P., Spano, G., Khomenko, I., Biasioli, F., Capozzi, V., Aznar, R. In situ riboflavin fortification of different kefir-like cereal-based beverages using selected Andean LAB strains. *Food microbiology* 2019, 77, 61-68.
DOI: <https://doi.org/10.1016/j.fm.2018.08.008>
6. Guarrasi, V., Sannino, C., Moschetti, M., Bonanno, A., Di Grigoli, A., Settanni, L. The individual contribution of starter and non-starter lactic acid bacteria to the volatile organic compound composition of Caciocavallo Palermitano cheese. *International journal of food microbiology* 2017, 259, 35-42.
DOI: <https://doi.org/10.1016/j.ijfoodmicro.2017.07.022>.
7. Mukisa, I. M., Byaruhanga, Y. B., Muyanja, C. M., Langsrud, T., Narvhus, J. A. Production of organic flavor compounds by dominant lactic acid bacteria and yeasts from Obushera, a traditional sorghum malt fermented beverage. *Food science & nutrition* 2017, 5(3), 702-712.
DOI: <https://doi.org/10.1002/fsn3.450>
8. Chai, L. J., Qiu, T., Lu, Z. M., Deng, Y. J., Zhang, X. J., Shi, J. S., Xu, Z. H. Modulating microbiota metabolism via bioaugmentation with *Lactobacillus casei* and *Acetobacter pasteurianus* to enhance acetoin accumulation during cereal vinegar fermentation. *Food Research International* 2020, 138, 109737.
DOI: <https://doi.org/10.1016/j.foodres.2020.109737>
9. Nielsen, G. S., Larsen, L. M., Poll, L. Formation of volatile compounds in model experiments with crude leek (*Allium ampeloprasum* Var. Lancelot) enzyme extract and linoleic acid or linolenic acid. *Journal of Agricultural and Food Chemistry* 2004; 52(8), 2315–2321.
DOI: <https://doi.org/10.1021/jf030600s>
10. Shimoda, M., Yoshimura, Y., Yoshimura, T., Noda, K., Osajima, Y. Volatile flavor compounds of sweetened condensed milk. *Journal of Food Science* 2001; 66(6), 804–807.
DOI: <https://doi.org/10.1111/j.1365-2621.2001.tb15176.x>
11. Fan, W., Qian, M. C. Characterization of aroma compounds of Chinese “Wuliangye” and “Jiannanchun” liquors by aroma extract dilution analysis. *Journal of Agricultural and Food Chemistry* 2006; 54(7), 2695–2704.
DOI: <https://doi.org/10.1021/jf052635t>
12. Ruiz, J., Antequera, T., Andres, A. I., Petron, M. J., Muriel, E. Improvement of a solid phase extraction method for analysis of lipid fractions in muscle foods. *Analytica Chimica Acta* 2004; 520, 201–205.
DOI: <https://doi.org/10.1016/j.aca.2004.04.059>
13. Salmerón, I., Rozada R., Thomas K., Ortega-Rivas E., and Pandiella S.S. Sensory characteristics and volatile composition of a cereal beverage fermented with *Bifidobacterium breve* NCIMB 702257. *Food Science and Technology International* 2014; 20: 205.
DOI: <https://doi.org/10.1177/1082013213481466>
14. Lee S.M., Lima H.J., Changb J.W., Hurhb B.S., Kima Y.K.; Investigation on the formations of volatile compounds, fatty acids, and ylactones in white and brown rice during fermentation. *Food Chemistry* 2018; 269: 347-354.
DOI: <https://doi.org/10.1016/j.foodchem.2018.07.037>
15. Ansorena, D., Gimeno, O., Astiasaran, I., Bello, J. Analysis of volatile compounds by GC-MS of a dry fermented sausage: Chorizo de Pamplona. *Food Research International* 2001, 34, 67–75.
DOI: [https://doi.org/10.1016/S0963-9969\(00\)00133-2](https://doi.org/10.1016/S0963-9969(00)00133-2)
16. Romero-Guido, C.; Belo, I.; Ta, T.M.N.; Cao-Hoang, L.; Alchihab, M.; Gomes, N.; Thonart, P.; Teixeira, J.A.; Destain, J.; Waché, Y. Biochemistry of lactone formation in yeast and fungi and its utilisation for the production of flavour and fragrance compounds. *Appl. Microbiol. Biotechnol.* 2011, 89, 535–547.
DOI: <https://doi.org/10.1007/s00253-010-2945-0>
17. Audrain, B., Farag, M. A., Ryu, C. M., & Ghigo, J. M. Role of bacterial volatile compounds in bacterial biology. *FEMS Microbiology Reviews* 2015, 39(2), 222–233
DOI: <https://doi.org/10.1093/femsre/fuu013>
18. Garrido-Galand S, Asensio-Grau A, Calvo-Lerma J, Heredia A, Andrés A. The potential of fermentation on nutritional and technological improvement of cereal and legume flours: A review. *Food Res Int.* 2021 Jul; 145:110398.
DOI: <https://doi.org/10.1016/j.foodres.2021>
19. Rodrigues, F.; Ludovico, P.; Leão, C. Sugar metabolism in yeasts: An overview of aerobic and anaerobic glucose catabolism. In *Biodiversity and Ecophysiology of Yeasts*; Edited by Péter, G., Rosa, C., Eds.; Springer: Berlin, Germany, 2006; pp. 101–121.
20. Vivíjs, B.; Moons, P.; Geeraerd, A.H.; Aertsen, A.; Michiels, C.W. 2,3-Butanediol fermentation promotes growth of *Serratia plymuthica* at low pH but not survival of extreme acid challenge. *Int. J. Food Microbiol.* 2014, 175, 36–44.
DOI: <https://doi.org/10.1016/j.ijfoodmicro.2014.01.017>
21. Pogačić, T.; Maillard, M.-B.; Leclerc, A.; Hervé, C.; Chuat, V.; Valence, F.; Thierry, A. *Lactobacillus* and *Leuconostoc* volatiles in cheese conditions. *Appl. Microbiol. Biotechnol.* 2016, 100, 2335–2346.
DOI: <https://doi.org/10.1007/s00253-015-7227-4>

22. Gonda, I.; Bar, E.B.; Portnoy, V.; Lev, S.; Burger, J.; Schaffer, A.A.; Tadmor, Y.; Gepstein, S.; Giovannoni, J.J.; Lewinsohn, N.K.E. Branched-chain and aromatic amino acid catabolism into aroma volatiles in Cucumis melo L. Fruit. *J. Exp. Bot.* 2010, 61, 1111–1123.
DOI: <https://doi.org/10.1093/jxb/erp390>
23. Landaud, S.; Helinck, S.; Bonnarme, P. Formation of volatile sulfur compounds and metabolism of methionine and other sulfur compounds in fermented food. *Appl. Microbiol. Biotechnol.* 2008, 77, 1191–1205.
DOI: <https://doi.org/10.1007/s00253-007-1288-y>
24. Yu, A.-Q.; Juwono, N.K.P.; Leong, S.S.J.; Chang, M.W. Production of fatty acid-derived valuable chemicals in synthetic microbes. *Front. Bioeng. Biotechnol.* 2014, 2, 78.
DOI: <https://doi.org/10.3389/fbioe.2014.00078>
25. Son, E.Y.; Lee, S.M.; Kim, M.J.; Seo, J.A.; Kim, Y.-S. Comparison of volatile and non-volatile metabolites in rice wine fermented by Koji inoculated with *Saccharomyces fibuligera* and *Aspergillus oryzae*. *Food Res. Int.* 2018, 109, 596–605.
DOI: <https://doi.org/10.1016/j.foodres.2018.05.008>
26. Hirata H, Ohnishi T, Watanabe N. Biosynthesis of floral scent 2-phenylethanol in rose flowers. *Biosci Biotechnol Biochem.* 2016:1865-73.
DOI: <https://doi.org/10.1080/09168451.2016.1191333>.
27. Dongmo, S. N., Sacher, B., Kollmannsberger, H., Becker, T. Key volatile aroma compounds of lactic acid fermented malt-based beverages—impact of lactic acid bacteria strains. *Food chemistry* 2017, 229, 565-573.
DOI: <https://doi.org/10.1016/j.foodchem.2017.02.091>
28. Dongmo, S. N., Sacher, B., Kollmannsberger, H., Becker, T. Key volatile aroma compounds of lactic acid fermented malt-based beverages—impact of lactic acid bacteria strains. *Food chemistry* 2017, 229, 565-573.
DOI: <https://doi.org/10.1016/j.foodchem.2017.02.091>
29. Zhao, J., Fleet, G. The effect of lactic acid bacteria on cocoa bean fermentation. *International Journal of food microbiology* 2015, 205, 54-67.
DOI: <https://doi.org/10.1016/j.ijfoodmicro.2015.03.031>
30. Mandha, J., Shumoy, H., Devaere, J., Schouteten, J. J., Gellynck, X., De Winne, A., Raes, K. Effect of lactic acid fermentation on volatile compounds and sensory characteristics of mango (*Mangifera indica*) juices. *Foods* 2022, 11(3), 383.
DOI: <https://doi.org/10.3390/foods11030383>
31. Ayseli, M. T., Ayseli, Y. İ. Flavors of the future: Health benefits of flavor precursors and volatile compounds in plant foods. *Trends in Food Science & Technology* 2016, 48, 69-77. DOI: <https://doi.org/10.1016/j.tifs.2015.11.005>
32. Antonio, A. G., Moraes, R. S., Perrone, D., Maia, L. C., Santos, K. R. N., Lório, N. L., Farah, A. Species, roasting degree and decaffeination influence the antibacterial activity of coffee against *Streptococcus mutans*. *Food Chemistry* 2010, 118(3), 782-788.
DOI: <https://doi.org/10.1016/j.foodchem.2009.05.063>
33. Flanagan, J., Bily, A., Rolland, Y., Roller, M. Lipolytic activity of Svetol®, a decaffeinated green coffee bean extract. *Phytotherapy research* 2014, 28(6), 946-948.
DOI: <https://doi.org/10.1002/ptr.5085>
34. Van Dam, R. M. Coffee and type 2 diabetes: from beans to beta-cells. *Nutrition, Metabolism and Cardiovascular Diseases* 2006, 16(1), 69-77.
DOI: <https://doi.org/10.1016/j.numecd.2005.10.003>
35. Younesi, E., Ayseli, M. T. An integrated systems-based model for substantiation of health claims in functional food development. *Trends in Food Science & Technology* 2015, 41(1), 95–100.
DOI: <https://doi.org/10.1016/j.tifs.2014.09.006>
36. Lanciotti, R., Gianotti, A., Patrignani, F., Belletti, N., Guerzoni, M. E., Gardini, F. Use of natural aroma compounds to improve shelf-life and safety of minimally processed fruits. *Trends in food science & technology* 2004, 15(3-4), 201-208. DOI: <https://doi.org/10.1016/j.tifs.2003.10.004>.
37. Park SH, Jo A, Lee KG. Effect of various roasting, extraction and drinking conditions on furan and 5-hydroxymethylfurfural levels in coffee. *Food Chem.* 2021 Oct 1; 358:129806.
DOI: <https://doi.org/10.1016/j.foodchem.2021.129806>.
38. Park, S.N., Lim, Y.K., Freire, M.O., Cho, E., Jin, D., Kook, J.K. Antimicrobial effect of linalool and α -terpineol against periodontopathic and cariogenic bacteria. *Anaerobe* 2012, 18(3), 369–372.
DOI: <https://doi.org/10.1016/j.anaerobe.2012.04.001>.
39. Kim A., Balanov P., Smotraeva I., Study of changes of antioxidant activity during fermentation of various types of legumes as a possible functional food ingredient. *Functional Foods in Health and Diseases* 2024, 751-776.
DOI: <https://doi.org/10.31989/ffhd.v14i11.1452>
40. Oi Y., Zhang W., Li N, Huang X., Lan J., A combination of multi-strain probiotics, prebiotic, and plant extracts improves ethanol-induced hangover outcomes in a zebrafish model *Functional Foods in Health and Diseases.* 2024, 728-738. DOI: <https://doi.org/10.31989/ffhd.v14i10.1462>
41. Mariod, A. A., Mustafa, E. M. A., Yahia, M. B. A review on the

42. health benefits of *Monechma ciliatum* (Black mahlab): A potential functional food. *Journal of Functional Foods in Health and Disease*, 2022 12(2).
DOI: <https://doi.org/10.31989/ffhd.v12i2.879>
43. Martínez-Mayorga, K., Franco, J.L.M. *Food informatics*. New York Dordrecht London: Springer Cham Heidelberg, (Chapter 3) 2014. 97-110 p.
44. Schwab, W., Davidovich-Rikanati, R., LewinsOhn, E. Biosynthesis of plant derived flavor compounds. *Plant Journal* 2008, 54(4), 712–732.
DOI: <https://doi.org/10.1111/j.1365-313X.2008.03446.x>
45. Lopez, H. L., Ziegenfuss, T. N., Hofheins, J. E., Habowski, S. M., Arent, S. M., Weir, J. P., Ferrando, A. Eight weeks of supplementation with a multi-ingredient weight loss product enhances body composition, reduces hip and waist girth, and increases energy levels in overweight men and women. *Journal of the International Society of Sports Nutrition* 2013, 10(22),1-14.
DOI: <https://doi.org/10.1186/1550-2783-10-22>.
46. Ayala-Zavala, J. F., González-Aguilar, G. a., Del-Toro-Sánchez, L. Enhancing safety and aroma appealing of fresh-cut fruits and vegetables using the antimicrobial and aromatic power of essential oils. *Journal of Food Science* 2009, 74(7), 84-90.
DOI: <https://doi.org/10.1111/j.1750-3841.2009.01294.x>
47. Keiler, A. M., Zierau, O., Kretzschmar, G. Hop extracts and hop substances in treatment of menopausal complaints. *Planta Medica* 2013, 79(7), 576–579.
DOI: <https://doi.org/10.1055/s-0032-1328330>
48. Rekha, K. R., Selvakumar, G. P., Sethupathy, S., Santha, K., Sivakamasundari, R. I. Geraniol ameliorates the motor behavior and neurotrophic factors inadequacy in MPTP-induced mice model of Parkinson's disease. *Journal of Molecular Neuroscience* 2013, 51(3), 851–862.
DOI: <https://doi.org/10.1007/s12031-013-0074-9>
49. Boulogne, I., Petit, P., Ozier-Lafontaine, H., Desfontaines, L., Loranger-Merciris, G. Insecticidal and antifungal chemicals produced by plants: A review. *Environmental Chemistry Letters* 2012, 10(4), 325–347.
DOI: <https://doi.org/10.1007/s10311-012-0359-1>
50. Hubert, J., Münzbergová, Z., Santino, A. Plant volatile aldehydes as natural insecticides against stored-product beetles. *Pest Management Science* 2008, 64, 57–64. Edited by Jeleń, H. *Food flavors chemical, sensory and technological properties*, New York: CRC Press, (Chapter 1) 2011.
51. Özçelik, B., Kartal, M., Orhan, I. Cytotoxicity, antiviral and antimicrobial activities of alkaloids, flavonoids, and phenolic acids. *Pharmaceutical Biology* 2011, 49(4), 396–402.
DOI: <https://doi.org/10.3109/13880209.2010.519390>
52. Fadida, T., Selilat-Weiss, A., Poverenov, E. N-hexylimine-chitosan, a biodegradable and covalently stabilized source of volatile, antimicrobial hexanal. Next generation controlled-release system. *Food Hydrocolloids* 2015, 48, 213–219.
DOI: <https://doi.org/10.1016/j.foodhyd.2015.02.033>
53. Li, Y., Lv, O., Zhou, F., Li, Q., Wu, Z., Zheng, Y. Linalool inhibits LPS induced inflammation in BV2 microglia cells by activating Nrf2. *Neurochemical Research* 2015, 40(7), 1520–1525. DOI: <https://doi.org/10.1007/s11064-015-1629-7>
54. Davidovich-Rikanati, R., Sitrit, Y., Tadmor, Y., Iijima, Y., Bilenko, N., Bar, E., Lewinsohn, E. Enrichment of tomato flavor by diversion of the early plastidial terpenoid pathway. *Nature Biotechnology* 2007, 25(8), 899–901.
DOI: <https://doi.org/10.1038/nbt1312>
55. Anuradha, K., Shyamala, B. N., Naidu, M.M. Vanilla-its science of cultivation, curing, chemistry, and nutraceutical properties. *Critical Reviews in Food Science and Nutrition* 2013, 53(12), 1250–76.
DOI: <https://doi.org/10.1080/10408398.2011.563879>.
56. Hannemann, A., Cytlak, U. M. C., Gbotosho, O. T., Rees, D. C., Tewari, S., Gibson, J. S. Effects of o-vanillin on K⁺ transport of red blood cells from patients with sickle cell disease. *Blood Cells, Molecules, and Diseases*, 2014 53(1-2), 21–26. DOI: <https://doi.org/10.1016/j.bcmd.2014.02.004>